Quick-Start Protocol May 2019

QlAamp® PowerFecal® Pro DNA Kit

Solution CD2 should be stored at 2–8°C. All other reagents and kit components should be stored at room temperature (15–25°C).

Further information

- QIAamp PowerFecal Pro DNA Kit Handbook: www.qiagen.com/HB-2560
- Safety Data Sheets: www.qiagen.com/safety
- Technical assistance: support.qiagen.com

Notes before starting

- Ensure that the PowerBead Pro Tubes rotate freely in the centrifuge without rubbing.
- If Solution CD3 has precipitated, heat at 60°C until precipitate dissolves.
- Perform all centrifugation steps at room temperature (15–25°C).
- Spin the PowerBead Pro Tube briefly to ensure that the beads have settled at the bottom.
 Add up to 250 mg of stool and 800 µl of Solution CD1. Vortex briefly to mix.
- 2. Secure the PowerBead Pro Tube horizontally on a Vortex Adapter for 1.5–2 ml tubes (cat. no. 13000-V1-24). Vortex at maximum speed for 10 min.

Note: If using the Vortex Adapter for more than 12 preps simultaneously, increase the vortexing time by 5–10 min.

Note: For more information about other bead beating methods, see the "Protocol: Detailed" section of QlAamp® PowerFecal® Pro DNA Kit Handbook.

- 3. Centrifuge the PowerBead Pro Tube at $15,000 \times g$ for 1 min.
- 4. Transfer the supernatant to a clean 2 ml Microcentrifuge Tube (provided).

Note: Expect 500–600 µl. The supernatant may still contain some stool particles.

- 5. Add 200 μl of Solution CD2 and vortex for 5 s.
- 6. Centrifuge at $15,000 \times g$ for 1 min at room temperature. Avoiding the pellet, transfer up to $700 \, \mu l$ of supernatant to a clean 2 ml Microcentrifuge Tube (provided).

Note: Expect 500–600 μl.

7. Add 600 µl of Solution CD3 and vortex for 5 s.



- 8. Load 650 μ l of the lysate onto an MB Spin Column and centrifuge at 15,000 x g for 1 min.
- 9. Discard the flow-through and repeat step 8 to ensure that all of the lysate has passed through the MB Spin Column.
- 10. Carefully place the MB Spin Column into a clean 2 ml Collection Tube (provided). Avoid splashing any flow-through onto the MB Spin Column.
- 11. Add 500 μ l of Solution EA to the MB Spin Column. Centrifuge at 15,000 x g for 1 min.
- 12. Discard the flow-through and place the MB Spin Column back into the same 2 ml Collection Tube.
- 13. Add 500 µl of Solution C5 to the MB Spin Column. Centrifuge at 15,000 x g for 1 min.
- 14. Discard the flow-through and place the MB Spin Column into a new 2 ml Collection Tube (provided).
- 15. Centrifuge at up to 16,000 x g for 2 min. Carefully place the MB Spin Column into a new 1.5 ml Elution Tube (provided).
- 16. Add 50–100 μ l of Solution C6 to the center of the white filter membrane.
- 17. Centrifuge at 15,000 x g for 1 min. Discard the MB Spin Column. The DNA is now ready for downstream applications.

Note: We recommend storing the DNA frozen ($-30 \text{ to } -15^{\circ}\text{C} \text{ or } -90 \text{ to } -65^{\circ}\text{C}$) as Solution C6 does not contain EDTA. To concentrate DNA, please refer to the Troubleshooting Guide.



Scan QR code for handbook.

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