Quick-Start Protocol February 2017

MagAttract® PowerClean® DNA Kit (384)

All reagents and kit components of the MagAttract PowerClean DNA Kit (384) should be stored at room temperature (15–25°C).

Further information

- Safety Data Sheets: **www.qiagen.com/safety**
- Technical assistance: **support.qiagen.com**

Notes before starting

- This kit was designed to work with up to 20 µg of input DNA and has been used successfully with as little as 20 ng. Using more than 20 µg will decrease the efficiency of inhibitor removal and can lead to loss of nucleic acid recovery.
- For best results, input DNA samples should be in water or Tris buffer, pH 8.0. We do not recommend using this kit with buffers containing > 1 mM EDTA.
- You will need 300 ml of 100% ethanol for each 96 well plate being processed on the KingFisher® Flex and 40 ml of 100% ethanol for each 12 wells being processed on the Kingfisher Duo.

1. Add up to 100 µl of DNA sample to each well of a 96 Well Plate (provided). If less than 100 µl is added, adjust the sample volume to 100 µl with molecular biology-grade water (user provided).
2. Add 50 µl of Solution CU to each well containing DNA. Mix by repeated pipetting (3X).
3. Add 50 µl of Solution IR to each well containing DNA. Seal the plate with a Sealing Tape and mix the contents in the wells by gently tapping on the sides of the sealed plate.
4. Centrifuge the plate at 4500 x *g* for 6 min at room temperature with an appropriately weighed balance plate.
5. To run the DNA purification on the KingFisher Flex, go to Step 6 (next page). If using the KingFisher Duo, please refer to the respective section of the Handbook.

KingFisher Flex protocol

6. Resuspend ClearMag® Beads (Zorb Reagent) by vortexing the bottle. For each 96 well plate to be processed, combine 45 ml of ClearMag Binding Solution and 2 ml of ClearMag Beads in a 50 ml conical tube (user provided). Mix well to obtain a homogeneous dispersion of beads.
7. Add 470 µl of the ClearMag Beads/Binding Solution to each well of a KingFisher Microtiter Deep Well 96-well plate.
8. Remove and discard the Sealing Tape from the plate centrifuged in Step 4. Avoiding the pellet, transfer the entire volume (expect 150–190 µl) of supernatant from each well in to a respective well in the KingFisher Microtiter Deep Well Plate prepared in Step 7.
   Note: We recommend that you use an 8-tip or 12-tip multichannel pipettor to reduce total processing time. You may find it helpful to place the 96 well plate on top of a colored (i.e., non-white) support to aid in visualizing the pellet at the bottom of each well.
9. Open the KingFisher Flex-specific PowerMag® DNA Clean-Up program on your instrument.
10. Place the KingFisher Microtiter Deep Well 96 Plate containing the DNA samples and ClearMag Beads/Binding Solution (from Step 8) onto the robotic deck at the specified location indicated in the PowerMag DNA Clean-Up program.
11. Place 1 ml of 100% ethanol (user provided) into each well of three clean KingFisher Microtiter Deep Well 96 plates. Place the pates on the deck at the specified locations indicated in the PowerMag DNA Clean-Up program.
12. Place 50–100 µl of Solution EB into each well of a KingFisher 96 KF Elution plate and place on the deck at the specified location. Initiate the KingFisher PowerMag DNA Clean-Up robotic program.
13. When the robotic program is complete, cover the wells of the KingFisher 96 KF Elution plate with an appropriate storage seal (user provided). The DNA is now ready for downstream applications.