

cador[®] Pathogen 96 QIAcube[®] HT Kit

This protocol is for use with QIAcube HT Prep Manager software. If you are using QIAcube HT 4.17 software, download the corresponding protocol at www.qiagen.com/HB-1569.

Further information

- *cador Pathogen 96 QIAcube HT Handbook*: www.qiagen.com/HB-2166
- Safety Data Sheets: www.qiagen.com/safety
- Technical assistance: support.qiagen.com

Notes before starting

- This protocol is for the purification of pathogen nucleic acids from various samples. See the *cador Pathogen QIAcube HT Handbook* for sample pretreatments.
 - Prepare Buffers ACB, AW1, AW2 and carrier RNA according to the instructions in the *cador Pathogen 96 QIAcube HT Handbook*.
 - Prepare a mixture of Buffer VXL, carrier RNA, proteinase K and internal control (if applicable) immediately before starting the run according to Table 1.
1. Start the QIAcube HT Prep Manager software. Click on the **Home** icon in the main toolbar to access the Home screen.
 2. Select *cador Pathogen 96* from the **Create Experiment** list. Follow the instructions in the wizard and fill in all required fields.
 3. In the **Setup** step, select **Sample type** and **Pre-treatment** for documentation.
 4. Select the protocol: *cador Pathogen* protocol (including lysis) or Heated off-board lysis protocol (without lysis). For information about optional steps and advanced options see the kit handbook.
 5. Define samples in the **Labware selection** step.

6. Arrange samples to the output plate in the **Assignment** step.

Note: The instrument must be switched on and connected to the software before entering the **Worktable** step.
7. Follow the instructions for loading the worktable.
8. Add the volume of sample indicated in the **Worktable** step to the selected S-Block wells.
9. Save the experiment by clicking the **Save** button in the button bar.
10. Click the **Start** run button to start the run.

Important: If the optional Vacuum performance check has been selected, the software will show a dialog that needs to be confirmed after defined vacuum steps.
11. When the protocol is complete, cover the elution plate (EMTR) with the lid and remove it from the elution chamber.

Note: If using Top Elute fluid, there may be 2 liquid phases in the elution microtubes. Top Elute fluid will be the top layer over the elution buffer.
12. Create a report (if required).
13. Follow the cleaning procedure.

Table 1. Buffer VXL mixture preparation.

Samples	24	32	40	48	56	64	72	80	88	96
Buffer VXL (ml)	4.6	5.9	7.4	8.8	9.9	11.8	12.7	14.1	15.5	17
Proteinase K (µl)	580	740	920	1100	1240	1470	1600	1800	2000	2200
Carrier RNA* (µl)	30	40	46	56	62	74	80	90	96	100
Internal Control* (µl)	280	370	460	550	620	730	800	880	980	1000

* If you are not using the IC or Carrier RNA then use RNase-free water instead.



Scan QR code for handbook.

For up-to-date licensing information and product-specific disclaimers, see the respective QIAGEN kit handbook or user manual.

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