

## Novel, Automated Procedure for Extraction of Microbial gDNA for High-Throughput Microbiome Analysis

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Novel, Procedure for Extraction of Microbial gDNA from Environmental Samples



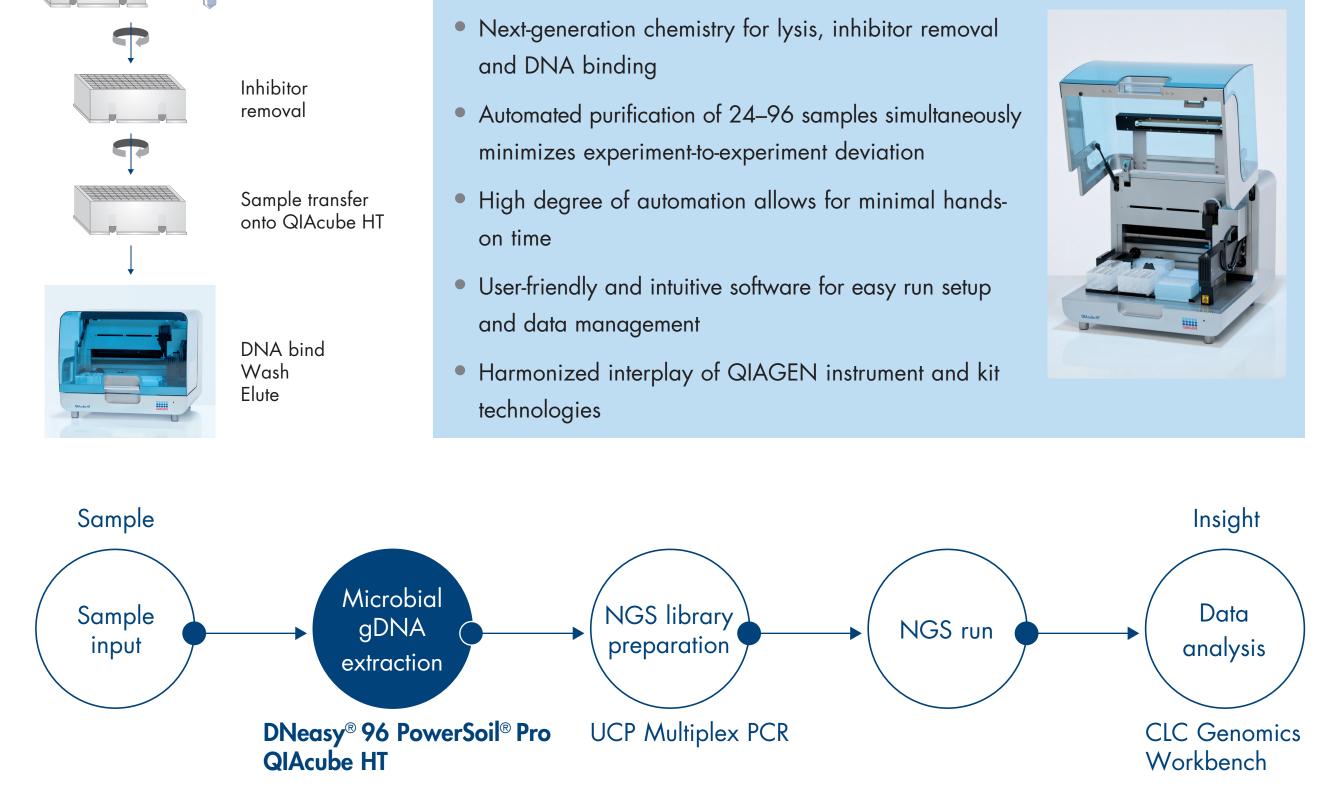


• New bead technology for efficient and unbiased lysis

Studies of the environmental and human microbiome face the problem that sample materials, such as soil and stool, are extremely difficult sample types. This is due to their high abundance of inhibitory substances such as humic and fulvic acids in soil, and bile acids and proteoglycans in stool. These inhibitory substances are often co-purified and have the potential to inhibit enzymatic reactions. Extraction of microbial DNA is therefore challenging and often requires tedious and time-consuming workflows, especially with large sample sizes in high-throughput studies.

To address these challenges, we have developed a second generation of the QIAGEN<sup>®</sup>/MO BIO Inhibitor Removal Technology<sup>®</sup>, which efficiently removes PCR inhibitors from even the most difficult soil types and stool samples. High-throughput bead-beating homogenization allows rapid, thorough, and unbiased lysis by a combination of mechanical and chemical methods. Inhibitory substances are removed by a dedicated precipitation step, all in a fast and streamlined workflow. Subsequent DNA purification is automated on a 96-well silica membrane plate format on the QIAcube® HT instrument using vacuum technology. This technology enables high-throughput processing of environmental samples for downstream applications, such as PCR amplification and NGS analysis, in under 3 hours.

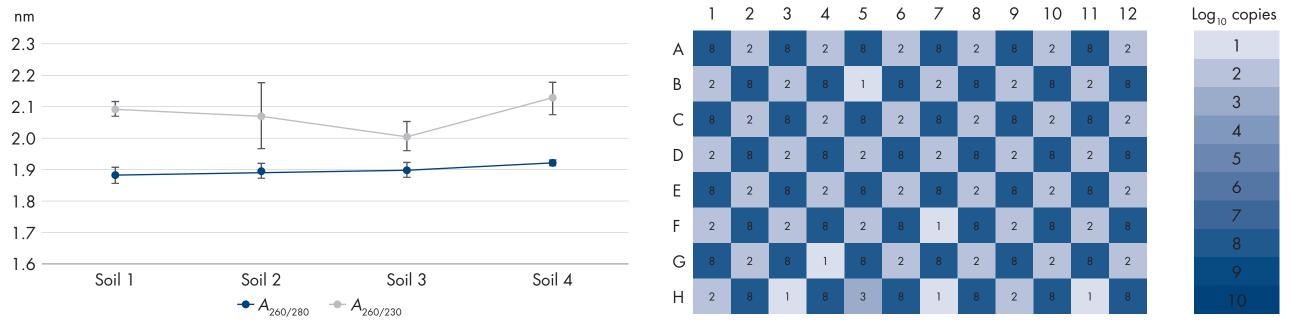
The automated protocol provides equivalent yields and purity to manual workflows, while enabling much higher throughput. 16S sequencing shows equivalent community composition to the manual methods, as shown by alpha and beta diversity measures. The new automated protocol allows for a complete picture of the diversity in the samples, thereby demonstrating that its suitability for high-throughput sample processing of both environmental and stool samples.

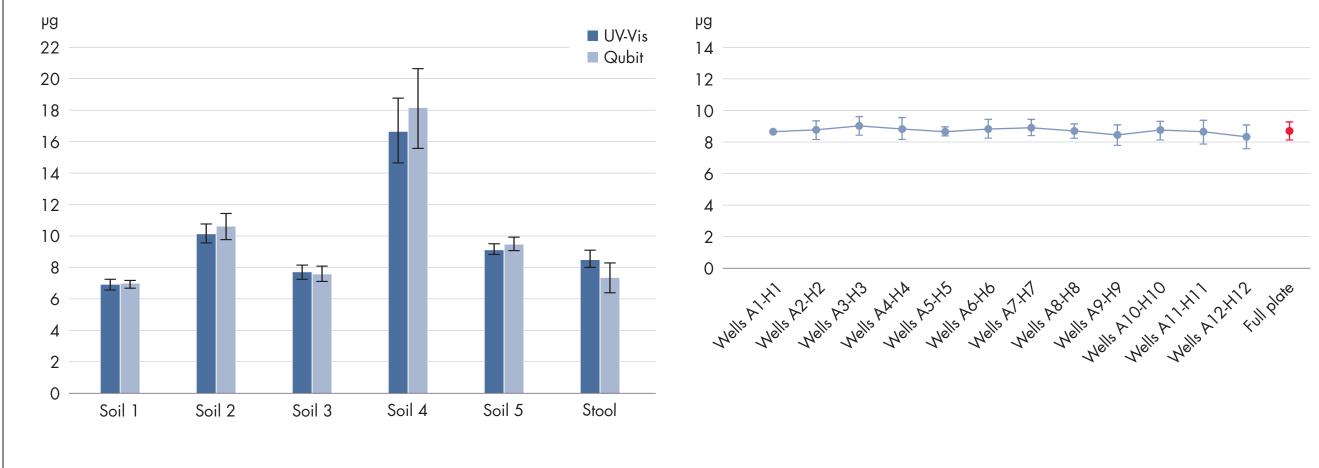


Reproducible, High-Quality DNA Yields from Soil and Stool

Soil and human stool samples were prepared using the new DNeasy 96 PowerSoil Pro QIAcube HT Kit. Microbial gDNA was either extracted from individually processed samples (A) or from aliquots of a lysate pool (B).

Inhibitor Removal Technology Increases Purity of Isolated DNA Soil samples were processed with the new DNeasy 96 PowerSoil Pro QIAcube HT Kit. 2 3 4 5 6 7 8 9 10 11 12 nm



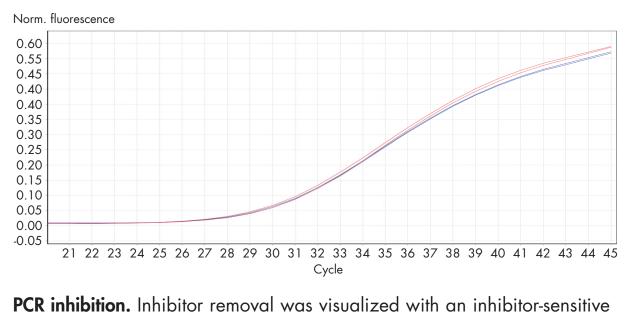


Yield from stool and soil. 250 mg of soil and 100 mg of human stool were prepared using the new DNeasy 96 PowerSoil Pro QIAcube HT Kit. Yields were measured by UV-Vis technology (NanoDrop<sup>™</sup>) and fluorometric quantification (Qubit<sup>®</sup>)

Consistency of yield across 96-well plates. DNA from a soil sample lysate pool was isolated in a full 96-well plate run on the QIAcube HT and yield determined by UV-Vis technology (NanoDrop).

The DNeasy 96 PowerSoil Pro QIAcube HT yields high amounts of DNA from soil and stool samples with high reproducibility and low variance over the 96-well plate.

 $A_{260/280}$  and  $A_{260/230}$  ratios. DNA purity shown as UV measurements and the analysis of ratios 260/280 (blue line) and 260/230 (grey line).



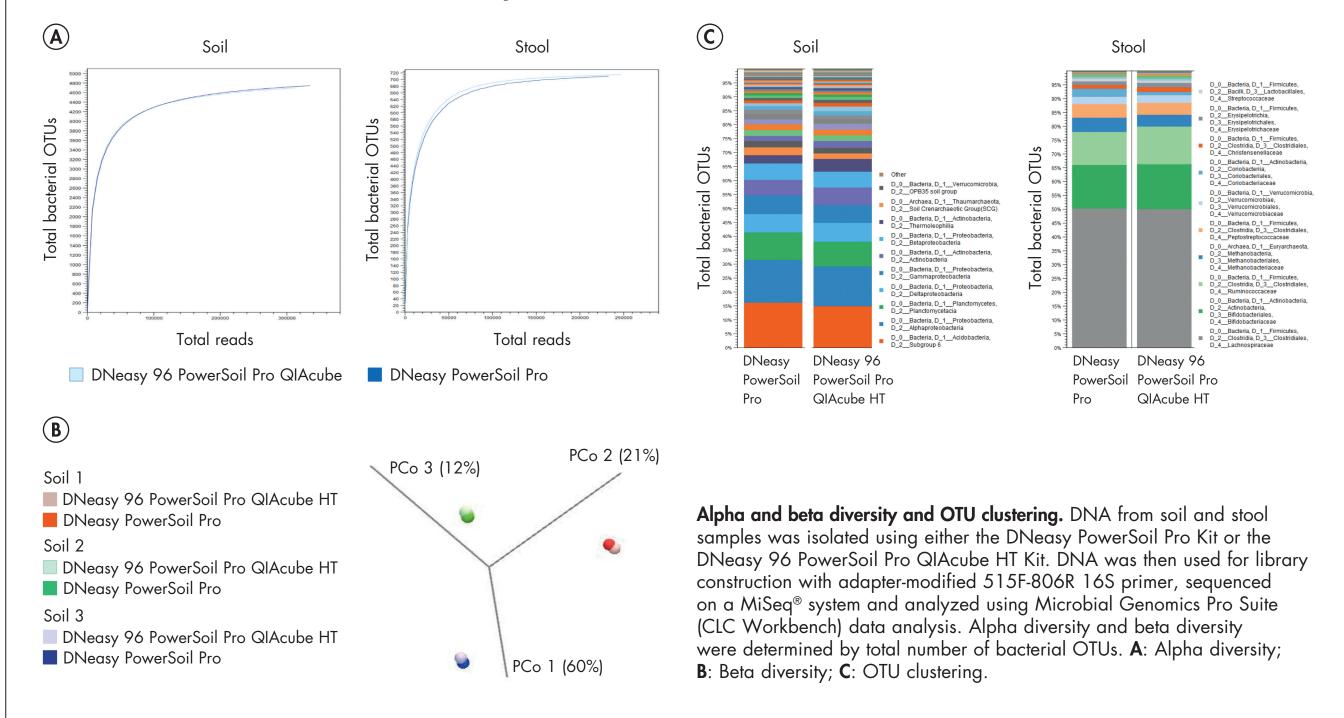
PCR with an internal control spiked with eluates from the new DNeasy 96 PowerSoil Pro QIAcube HT Kit (blue lines). Eluate-free PCR reactions served as controls (red lines).

**Cross-contamination.** Aliquots from a soil sample lysate pool and H<sub>2</sub>O as substitute sample were distributed in a 96-well plate in a checkerboard pattern and processed on the QIAcube HT using the DNeasy 96 PowerSoil Pro protocol. Bacterial gDNA was determined by 16S qPCR and DNA content per well pictured by color intensity of blue. Dark blue: Soil samples with up to  $3.9 \times 10^8$  copies/well; Light blue: H<sub>2</sub>O samples with <100 copies/well.

DNeasy 96 PowerSoil Pro QIAcube HT shows: • A<sub>260</sub>/A<sub>280</sub> ratios ~1.9 • High  $A_{260}/A_{230}$  ratios over 2 • No inhibition in PCR

• Well controlled cross-contamination

Identification of Total Diversity and Community Representation in Soil and Stool Samples



## Conclusions

Relatively inexpensive, highly multiplexed 16S sequencing enables studies of large sample size, for which improved sample extraction techniques are needed in order to enable true high-throughput studies. A new semi-automated, high-throughput method is presented that:

- Allows simultaneous purification of microbial genomic DNA of up to 96 samples in a convenient semi-automated workflow using the QIAcube HT instrument
- Efficiently lyses bacteria in soil and stool samples
- Provides high yields of useful, pure DNA

Total bacterial OTUs identified in DNA isolated with the DNeasy 96 PowerSoil Pro QIAcube HT Kit and the DNeasy PowerSoil Pro Kit are comparable, showing an equally high alpha diversity and identical beta diversity.

• Removes inhibitors commonly present in soil and stool samples, allowing eluates to be used directly in downstream NGS applications.

These data show that the new high-throughput method on the QIAcube HT achieves successful DNA isolation from soil and stool samples in a semi-automated 96-well format.

The method presented here is intended for molecular biology applications. This method is not intended for the diagnosis, prevention, or treatment of a disease.

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