



March 2024

DNeasy[®] Plant Handbook

DNeasy Plant Mini Kit

DNeasy Plant Maxi Kit

For purification of total cellular DNA from plant cells and tissues or fungi

DNeasy 96 Plant Kit

For high-throughput purification of DNA from tissue

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Kit Contents

DNeasy Plant Kits	Mini (50)	Mini (250)	Maxi (24)
Catalog no.	69104	69106	68163
Number of preps	50	250	24
DNeasy Mini Spin Columns (colorless)	50	250	–
DNeasy Maxi Spin Columns (colorless)	–	–	24
QIAshredder Mini Spin Columns (lilac)	50	250	–
QIAshredder Maxi Spin Columns (lilac)	–	–	24
Collection tubes (2 mL)	50	250	–
Collection tubes (50 mL)	–	–	24
Buffer AP1	40 mL	200 mL	140 mL
Buffer P3	20 mL	2 x 50 mL	50 mL
Buffer AW1 (concentrate)*†	2 x 19 mL	151 mL	151 mL
Buffer AW2 (concentrate)†	17 mL	2 x 40 mL	2 x 68 mL
Buffer AE	2 x 12 mL	2 x 60 mL	60 mL
RNase A (100 mg/mL)	220 µL	5 x 220 µL	440 µL
Quick-Start Protocol	1	1	1

* Contains a chaotropic salt. Not compatible with disinfectants containing bleach. See “Safety Information”.

† Buffer AW1 and Buffer AW2 are supplied as concentrates. Add ethanol (96–100%) according to the bottle label before use to obtain a working solution.

DNeasy 96 Plant Kits	(6)
Catalog no.	69181
No. of preps	6
DNeasy 96 Plates	6
S-Blocks*	2
Collection microtubes, 1.2 mL (racked)	12 x 96
Collection microtube caps	4 x (120 x 8)
Elution Microtubes RS (racked) and caps	6 x 96
AirPore Tape Sheets	1 x 5 + 1 x 25
Buffer AP1	2 x 140 mL
Buffer P3	2 x 50 mL
Buffer AW1 (concentrate) ^{††}	151 mL
Buffer AW2 (concentrate) ^{††}	2 x 81 mL
Buffer AE	128 mL
RNase A (100 mg/mL)	2 x 440 µL
Reagent DX	1 mL
96-well-plate registers	8
Quick-Start Protocol	1

* Reusable; see “Appendix B: Recovery and Cleaning of Beads and S-Blocks” for cleaning instructions.

† Contains a chaotropic salt. Not compatible with disinfectants containing bleach. See “Safety Information”.

†† Buffer AW1 and Buffer AW2 are supplied as concentrates. Add ethanol (96–100%) according to the bottle label before use to obtain a working solution.

Shipping and Storage

Upon arrival, store the DNeasy Maxi Spin Columns at 2–8°C. All other components of DNeasy Plant Kits, including RNase A stock solution, should be stored dry, at room temperature (15–25°C), and are stable for 1 year under these conditions, if not otherwise stated on label.

For storage longer than 1 year or if ambient temperatures often exceed 25°C, we recommend keeping the RNase A stock solution at 2–8°C.

Intended Use

DNeasy Plant Kits are intended for molecular biology applications. These products are not intended for the diagnosis, prevention, or treatment of a disease.

QIAcube® Connect is designed to perform fully automated purification of nucleic acids and proteins in molecular biology applications. The system is intended for use by professional users trained in molecular biological techniques and the operation of QIAcube Connect.

All due care and attention should be exercised in the handling of the products. We recommend all users of QIAGEN® products to adhere to the NIH guidelines that have been developed for recombinant DNA experiments, or to other applicable guidelines.

Safety Information

When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, please consult the appropriate safety data sheets (SDSs). These are available online in convenient and compact PDF format at www.qiagen.com/safety, where you can find, view, and print the SDS for each QIAGEN kit and kit component.

<p>CAUTION</p> 	<p>DO NOT add bleach or acidic solutions directly to waste containing Buffer AW1</p>
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Buffer AW1 contains guanidine salts, which can form highly reactive compounds when combined with bleach. If liquid containing these buffers is spilt, clean with a suitable laboratory detergent and water. If the spilt liquid contains potentially infectious agents, clean the affected area first with laboratory detergent and water, and then with 1% (v/v) sodium hypochlorite.

Quality Control

In accordance with QIAGEN's ISO-certified Quality Management System, each lot of DNeasy Plant Kit is tested against predetermined specifications to ensure consistent product quality.

Introduction

DNeasy Plant Kits provide a fast and easy way to purify DNA from plant and fungal tissue. Up to 100 mg of tissue can be processed using the DNeasy Plant Mini Kit or up to 1 g of tissue using the DNeasy Plant Maxi Kit. The DNeasy 96 Kit is designed for high-throughput DNA purification from 50 mg plant tissue per well (for some plant tissues, up to 100 mg per well can be used).

Easy-to-use DNeasy Plant procedures provide pure total DNA (genomic, mitochondrial, and chloroplast) for reliable PCR and Southern blotting in less than 1 hour (DNeasy Plant Mini Kit) or less than 2 hours (DNeasy Plant Maxi Kit or DNeasy 96 Plant Kit). Purification requires no phenol or chloroform extraction or alcohol precipitation and involves minimal handling. This makes DNeasy Mini and Maxi Kits highly suited for simultaneous processing of multiple samples. For higher-throughput applications, the DNeasy 96 Plant Kit enables simultaneous processing of 96 or 192 samples.

Purified DNA is eluted in low-salt buffer or water, ready for use in downstream applications. DNA purified using DNeasy Plant Kits is up to 40 kb in size, with fragments of 20–25 kb predominating. DNA of this length denatures completely in PCR and shows the highest amplification efficiency. Purified DNA has an A_{260}/A_{280} ratio of 1.7–1.9, indicating high purity of the DNA.

The DNeasy membrane ensures complete removal of all inhibitors of PCR and other enzymatic reactions. DNA purified using DNeasy Plant Kits is highly suited for use in all downstream applications, including PCR, random amplified polymorphic DNA (RAPD) analysis, amplified fragment length polymorphism (AFLP) analysis, restriction fragment length polymorphism (RFLP) analysis, Southern blotting, microsatellite analysis, single nucleotide polymorphisms (SNP) genotyping, and quantitative real-time PCR.

The use of TissueRuptor® II or TissueLyser, for rapid and convenient disruption of plant tissue samples, is recommended for the most efficient processing in DNeasy Plant procedures.

Principle and procedure

Disruption of plant tissue

Complete and quick disruption of starting material is essential to ensure high DNA yields and to avoid DNA degradation. DNeasy Plant procedures are optimized for use with leaf tissues but can also be used to purify DNA from other plant tissues and fungi, including seeds and spores. However, when using tissues other than leaves, the disruption method may require optimization to ensure maximum DNA yield and quality.

DNeasy Plant Kits are compatible with all sample-disruption methods. Optimal results are obtained using the TissueRuptor II homogenizer or the TissueLyser system.

TissueRuptor II homogenizer

The TissueRuptor II is a hand-held rotor–stator homogenizer designed for rapid, efficient, and flexible disruption of a variety of biological samples, including plant and animal tissues. Samples are simultaneously disrupted and homogenized by a rotating blade (rotor–stator homogenization). TissueRuptor disposable probes enable flexible sample disruption in a wide range of volumes and formats. Using a separate disposable probe for each sample provides ease of use and prevents cross-contamination.

Fresh, frozen, or lyophilized plant tissue samples can be processed using the TissueRuptor II. Tissues can be disrupted in liquid nitrogen or directly disrupted in lysis buffer without using liquid nitrogen, depending on the protocol and downstream application used. See “Disruption and homogenization using the TissueRuptor II” for further details.

TissueLyser system

The TissueLyser system provides convenient, simultaneous disruption, and homogenization of multiple biological samples through high-speed shaking in plastic tubes with stainless steel, tungsten carbide, or glass beads. The TissueLyser III (cat. no. 9003240) provides medium- to high-throughput sample disruption of up to 48 or 192 samples. The TissueLyser LT (cat. no. 85600) provides fast low- to medium-throughput sample disruption of up to 12 samples.

Fresh, frozen, or lyophilized plant tissue samples can be processed using the TissueLyser. Fresh material can be directly disrupted in lysis buffer without using liquid nitrogen. Alternatively, fresh or frozen samples can be disrupted after freezing in liquid nitrogen without lysis buffer. Lyophilized tissue can be disrupted without buffer at ambient temperature. See “Disruption and homogenization using the TissueLyser system” for further details.

DNA purification

In DNeasy Plant procedures (page 11), plant material is first mechanically disrupted and then lysed by addition of lysis buffer and incubation. RNase A in the lysis buffer digests the RNA in the sample. After lysis, proteins and polysaccharides are salt-precipitated.

In the DNeasy 96 procedures, cell debris and precipitates are removed by centrifugation. In DNeasy Mini and Maxi procedures, cell debris and precipitates are removed in a single step by a brief spin through the QIAshredder column, a unique filtration and homogenization unit. Binding buffer and ethanol are added to the cleared lysate to promote binding of the DNA to the DNeasy membrane. The sample is then applied to a DNeasy spin column or DNeasy 96 plate and then centrifuged. DNA binds to the membrane, while contaminants such as proteins and polysaccharides are efficiently removed by 2 wash steps. Pure DNA is eluted in a small volume of low-salt buffer or water. DNeasy-purified DNA has A_{260}/A_{280} ratios of 1.7–1.9, and absorbance scans show a symmetric peak at 260 nm, confirming high purity.

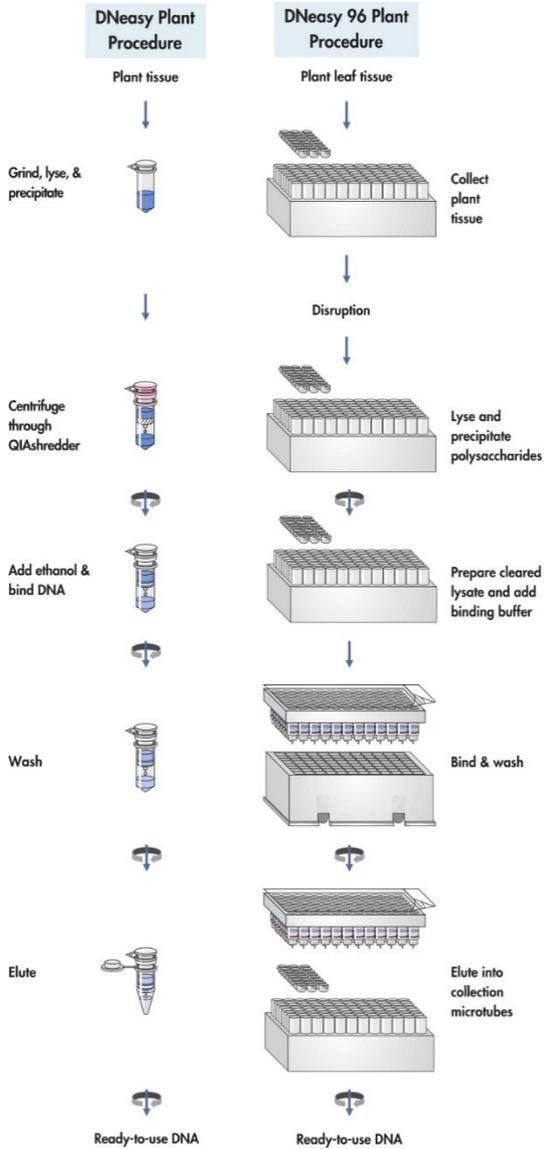


Figure 1. DNeasy Plant Procedure.

The DNeasy membrane combines the binding properties of a silica-based membrane with simple microspin technology or with the QIAGEN 96-Well-Plate Centrifugation System. DNA adsorbs to the DNeasy membrane in the presence of high concentrations of chaotropic salt, which remove water from hydrated molecules in solution. Buffer conditions in DNeasy Plant procedures are designed to enable the specific adsorption of DNA to the silica membrane and the optimal removal of carbohydrates, polyphenolics, and other plant metabolites.

Description of protocols

Different protocols in this handbook provide detailed instructions for using DNeasy Kits in the purification of genomic DNA from plant tissue.

“Protocol: Purification of Total DNA from Plant Tissue (Mini Protocol)” is for use with the DNeasy Plant Mini Kit, with up to 100 mg (wet weight) fresh, frozen, or lyophilized plant tissue, following the specifications in Table 1, next page.

“Protocol: Purification of Total DNA from Plant Tissue (Maxi Protocol)” is for use with the DNeasy Plant Maxi Kit, with up to 1 g (wet weight) fresh, frozen, or lyophilized plant tissue, following the specifications in Table 1.

“Protocol: Purification of Total DNA from Fresh Plant Tissue (DNeasy 96 Protocol)” is for use with the DNeasy 96 Plant Kit, with fresh plant tissue, following the specifications in Table 1. This protocol is optimized for use with the TissueLyser system for efficient and convenient high-throughput disruption of plant tissue.

“Protocol: Purification of Total DNA from Frozen or Lyophilized Plant Tissue (DNeasy 96 Protocol)” is for use with the DNeasy 96 Plant Kit, with frozen or lyophilized plant tissue, following the specifications in Table 1. This protocol is optimized for use with the TissueLyser system for efficient and convenient high-throughput disruption of plant tissue.

Table 1. Specifications for DNeasy Plant Kits

DNeasy Plant Kit	Mini	Maxi	96 (amount per well)
Maximum amount of starting material*	100 mg wet weight 20 mg dry weight	1 g wet weight 0.2 g dry weight	50 mg wet weight 10 mg dry weight
DNA-binding capacity†	50 µg	500 µg	50 µg
Maximum loading volume	700 µL	1.5 mL	1 mL
Elution volume	50–400 µL	500–2000 µL	100–200 µL
DNA-binding capacity†	50 µg	500 µg	50 µg
Typical yields	3–30 µg	30–260 µg	1–15 µg
Preparation time	<1 hour	<2 hours	<2 hours (192 samples)

* Exceeding the recommended amount of starting material will reduce yield and purity.

† Typically, the binding capacity will not be reached, because the DNA content of the recommended amounts of starting material will not exceed 30 µg (Mini), 300 µg (Maxi), or 50 µg (96).

Automated purification of DNA on QIAcube Instruments

Purification of DNA can be fully automated on QIAcube Connect or the classic QIAcube. The innovative QIAcube instruments use advanced technology to process QIAGEN spin columns, enabling seamLess integration of automated, low-throughput sample prep into your laboratory workflow. Sample preparation using QIAcube instruments follows the same steps as the manual procedure (i.e., lyse, bind, wash, and elute), enabling you to continue using the DNeasy Plant Kit for purification of high-quality DNA.

QIAcube instruments are preinstalled with protocols for purification of plasmid DNA, genomic DNA, RNA, viral nucleic acids and proteins, plus DNA and RNA cleanup. The range of protocols available is continually expanding, and additional QIAGEN protocols can be downloaded free of charge at www.qiagen.com/qiacubeprotocols



Figure 2. QIAcube Connect.

Equipment and Reagents to be Supplied by User

When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate safety data sheets (SDSs), available from the product supplier.

For all protocols

- Equipment for disruption and homogenization
We recommend either using the TissueLyser with the accessories listed in Table 2 or using the TissueRuptor II.

Table 2. TissueLyser accessories

Protocol	TissueLyser accessories
DNeasy Mini	Adaptor set 2 x 24 (cat. no. 69982) Tungsten carbide beads, 3 mm (cat. no. 69997)* 1.5 mL or 2 mL safe-lock microcentrifuge tubes
DNeasy Maxi	Grinding jar set, stainless steel (cat. no. 69985)
DNeasy 96	Adaptor set 2 x 96 (cat. no. 69984) Tungsten carbide beads, 3 mm (cat. no. 69997)* Collection microtubes (included with DNeasy 96 Plant Kit)

* Stainless steel beads, 5 mm (cat. no. 69989), can also be used. We recommend the use of tungsten carbide beads because these perform better and more consistently than stainless steel beads.

- Pipettes and pipette tips
- Water bath or heating block for heating at 65°C and 80°C
- Vortexer
- Ethanol (96–100%)*
- Liquid nitrogen

* Do not use denatured alcohol, which contains other substances such as methanol or methylethylketone.

Note: Not required when processing lyophilized plant material or when processing fresh plant material in the “Protocol: Purification of Total DNA from Fresh Plant Tissue (DNeasy 96 Protocol)”.

For DNeasy Mini protocol

- Microcentrifuge tubes (1.5 mL)
- Microcentrifuge with rotor for 2 mL tubes
- Ice

For DNeasy Maxi protocol

- 15 mL and 50 mL centrifuge tubes
The use of disposable polypropylene tubes is recommended. Tubes used for the lysis step should be capable of withstanding the g-forces involved in centrifugation and should also be compatible with liquid nitrogen.
- Laboratory centrifuge (capable of 3000–5000 x *g*) equipped with a swing-out rotor
- All centrifugation steps are carried out in a conventional laboratory centrifuge (e.g., SIGMA® 6-16 series from SIGMA Laborzentrifugen GmbH, www.sigma-zentrifugen.de)* using a swinging bucket rotor. DNeasy Maxi spin columns and QIAshredder Maxi spin columns fit into the 50 mL centrifuge tubes provided. These tubes are compatible with almost all laboratory centrifuges and rotors. In the unlikely event that these tubes do not fit your rotor, the spin columns can also be used with any other commercially available 50 mL polypropylene or glass tubes.
- Spatula
- Ice

* This is not a complete list of suppliers and does not include many important vendors of biological supplies.

For DNeasy 96 protocol

- Centrifuge 4 16S or 4 16KS with Plate Rotor 2 x 96 (see page 23)
- Multichannel pipette with extended tips
- For efficient processing, we recommend the use of an electric multichannel pipette with a capacity of at least 1 mL per pipette tip. We recommend using extended tips with a maximum volume of 1250 μ L.
- Reagent reservoirs for multichannel pipettes
- Freezer or cold room at -30°C to -15°C

Important Notes

Collection and storage of starting material

After harvesting, if plant tissue will not be used immediately, it should be frozen in liquid nitrogen. It can then be stored at -90°C to -65°C for later processing. Ground tissue powder can also be stored at -90°C to -65°C . Alternatively, tissue can be dried or lyophilized after harvesting to allow storage at room temperature. To ensure DNA quality, samples should be completely dried within 24 hours of collection.

If possible, it is preferable to collect young materials (e.g., leaves, needles) because they contain more cells per weight and therefore result in higher yields. In addition, young leaves and needles contain smaller amounts of polysaccharides and polyphenolics and are therefore easier to handle.

When working with fungi, harvest mycelium directly from a culture dish or from liquid culture. For liquid culture, start by pelleting cells using centrifugation. Remove the supernatant completely before disruption and lysis. Fresh, frozen, or freeze-dried fungal material can be used.

Sample size

DNeasy Plant procedures are optimized for a maximum of 100 mg (Mini), 1 g (Maxi), or 50 mg (DNeasy 96) of wet-weight starting material.

Table 3 (next page) provides guidelines for wet weights of leaf tissue. If using dried starting material, the maximum amount that can be processed must be reduced by a factor of approximately 5. Exceeding the recommended maximum amount of starting material will result in inefficient lysis, resulting in low yield and purity.

With some plant species, it may be possible to increase the amount of starting material in the DNeasy 96 procedure to increase DNA yield. For example, increasing the amount of wheat starting material to 100 mg increases the DNA yield by 35% (27.4 µg DNA compared with 20.4 µg DNA, after two 100 µL elutions). Note that DNA yields do not necessarily increase linearly with increased amounts of starting material. Furthermore, for some plant species, impurities may be present in the purified DNA if the amount of starting material is increased. To find the optimum amount of starting material for a particular plant species, use a range of amounts (e.g., 50, 75, and 100 mg), and then determine the amount that provides the highest DNA yield and purity.

Note: The maximum amount of starting material that can be used with the collection microtubes provided in the DNeasy 96 Plant Kit is 50 mg (wet weight). For more than 50 mg starting material, other disruption vessels should be used.

Table 3. Approximate wet weights of leaf material

Sample	Size	Approximate wet weight (mg)
Leaf (punch)	1.5 cm diameter	25–75
Leaf (surface area)	12 cm ² (e.g., 4 x 3 cm)	170–510

Note: DNA yields vary depending on genome size, ploidy, and age of sample. Yields typically range from 3–30 µg per 100 mg of wet-weight sample.

Disruption and homogenization using the TissueRuptor II

Homogenization using the TissueRuptor II system (Figure 3, page 21) is carried out by first installing the adapter on the motor drive and placing a disposable probe into the adapter (see the *TissueRuptor II User Manual*). Fresh or frozen samples can be disrupted after freezing in liquid nitrogen without Buffer AP1. Alternatively, fresh material can be directly disrupted in Buffer AP1 without using liquid nitrogen, but this may cause shearing of high-molecular-weight DNA. We do not recommend disrupting frozen material in lysis buffer, because this can result in low yields and degraded DNA.

Especially hard tissues (such as roots or seeds) could cause the disposable probes to break and may not be well-suited for use with the TissueRuptor II. These tissues can be disrupted using a mortar and pestle or the TissueLyser system.

For optimal results, we recommend keeping the disruption time as short as possible. Disruption for more than 1 minute may lead to shearing of genomic DNA.

Note: Disruption can be performed without lysis buffer by keeping the sample submerged in liquid nitrogen before and during disruption. In this case, Buffer AP1 and RNase A stock solution (100 mg/mL) must be added to the sample immediately after disruption.

Disruption and homogenization using the TissueLyser system

Fresh, frozen, or lyophilized plant tissue samples can be processed using the TissueLyser system, including the TissueLyser II and the TissueLyser LT (Figure 3, page 21). Fresh material can be directly disrupted in lysis buffer without using liquid nitrogen. Alternatively, fresh or frozen samples can be disrupted after freezing the collection microtubes including the material on dry ice without lysis buffer. Freezing the racked collection microtubes in liquid nitrogen can cause breakage of the tubes. Lyophilized tissue can be disrupted without buffer at ambient temperature. Disruption of samples in lysis buffer yields DNA ideal for PCR, while disruption of samples in liquid nitrogen yields DNA of a higher molecular weight. We do not recommend disrupting frozen material in lysis buffer; this results in low yields and degraded DNA.

Important: When using the TissueLyser with frozen plant material, do not allow the adaptor plates to come into contact with liquid nitrogen.

Note: When using the TissueLyser Adapter Sets, samples on the inside of the adaptor rack move more slowly than samples on the outside. To prevent variation in sample homogenization, the adaptor sets should be removed from the TissueLyser and disassembled after the first disruption step. For the second disruption step, the adaptor sets should be reassembled so that the samples tubes that were outermost in the rack are

now innermost. Rearranging the racks of collection microtubes in this way ensures that all samples are thoroughly and equally disrupted.

Important: Merely rotating the entire plate sandwich so that the QIAGEN logos are upside down when reinserted into the TissueLyser is not sufficient, because the same samples that were outermost during the initial disruption will remain outermost in the second disruption step.

An instruction manual is provided with the TissueLyser. Read this manual carefully before using the TissueLyser. Do not use the TissueLyser with equipment other than that supplied.



Figure 3. The TissueLyser II, TissueLyser LT, and TissueRuptor II for sample disruption.

DNeasy Mini procedure

Plant material and a tungsten carbide bead are added to a 2 mL safe-lock tube. Tubes are placed into the adaptor sets, which are fixed into the clamps of the TissueLyser. Disruption is performed in two 1–2 minute high-speed (20–30 Hz) shaking steps. The beads are reusable (see “Appendix B: Recovery and Cleaning of Beads and S-Blocks” for cleaning instructions).

For optimal operation, the TissueLyser should always be balanced. A balance can be provided by assembling a second adaptor using safe-lock tubes without samples or buffers but containing the beads, and fixing this second adaptor into the empty clamp.

DNeasy Maxi procedure

Plant material is added to the grinding jars. The grinding balls are added, and the jars are fixed in the clamps of the TissueLyser. Disruption is performed in two 1–2 minute high-speed (20–30 Hz) shaking steps. The grinding jars and balls are reusable (see “Appendix B: Recovery and Cleaning of Beads and S-Blocks” for cleaning instructions).

For optimal operation, the TissueLyser should always be balanced. If using a single grinding jar, the balance should consist of a second grinding jar containing a stainless steel ball.

DNeasy 96 procedures

For DNeasy 96 procedures, plant material and a tungsten carbide bead are added to each of the 192 collection microtubes in 2 racks. The racks are placed into the adaptor sets, which are fixed into the clamps of the TissueLyser. Disruption is performed in two 1–2 minute high-speed (20–30 Hz) shaking steps. The beads are reusable (see “Appendix B: Recovery and Cleaning of Beads and S-Blocks” for cleaning instructions).

For optimal operation, the TissueLyser should always be balanced. A balance can be provided by assembling a second adaptor using a rack of collection microtubes without samples or buffers but containing the beads, and fixing this second adaptor into the empty clamp.

Centrifugation (DNeasy 96 procedures)

Centrifuges 4-16S or 4-16KS

DNeasy 96 spin protocols use a streamlined centrifugation procedure that enables purification of DNA from as many as 2 x 96 samples, in parallel, for direct use in any downstream application. The DNeasy 96 Plant procedure requires the use of QIAGEN 96 Well Plate Centrifugation System, comprising the Plate Rotor 2 x 96 and either the tabletop Centrifuge 4 16S or the refrigerated table-top Centrifuge 4 16KS. In addition to the Plate Rotor 2 x 96, a wide range of other rotors can be used with these centrifuges.

Standard table-top centrifuges and microtiter plate rotors are not suitable for the DNeasy 96 protocol for 2 reasons:

- The microtiter plate buckets are either not deep enough for the complete DNeasy 96 package or they will not swing out properly.
- High g-forces (>5500 x g) are required for optimal performance of the DNeasy 96 procedure. The speed limit of the Centrifuge 4 16S and the Centrifuge 4 16KS (6000 rpm; 5796 x g) is programmed so that the given g-force will not be exceeded.

All centrifugation steps are performed at room temperature.

Important: Centrifuges must be properly maintained for optimal performance. It is particularly important that the buckets and rotor pins are routinely greased to prevent suboptimal running conditions that may lead to cracking of DNeasy 96 plates.

For further information about QIAGEN Centrifuges and the Plate Rotor 2 x 96, contact QIAGEN Technical Services or your local distributor.

Note: If the Centrifuge 4 16KS is used, set the temperature to 40°C for all centrifugation steps.

Note: Use AirPore Tape Sheets (provided) to seal DNeasy 96 plates during all centrifugation steps to prevent cross-contamination between samples.

Abbreviated instructions for using the Centrifuge 4 16S and Centrifuge 4 16KS

Warning: Never run the centrifuge with empty plate carriers placed inside the buckets, that is, without the collection microtubes or DNeasy 96 plates and S-Blocks. If unsupported, the carriers will collapse under high g-forces. Therefore, remove the carriers during test runs. Standard microtiter plates may be centrifuged in the same carriers if the g-force does not exceed 500 x *g*.

Procedure

1. Switch on the centrifuge by pressing the main switch at the back.
2. Select the rotor selection list in the display field by turning the knob. After pressing the knob, turn the knob again to select the rotor/bucket combination "09100/09158" for the Plate Rotor 2 x 96. Confirm entry by pressing the knob. Entering the rotor number automatically sets the time and speed limits for centrifugation for that particular rotor, thus eliminating the danger of the centrifuge over-speeding.
3. Select **Speed** by turning the knob. Press the knob, and then, by turning the knob again, set the speed to **6000**. Confirm entry by pressing the knob. The corresponding relative centrifugal force (RCF) is calculated from the rotor number and speed and appears automatically in the RCF field. It is also possible to enter the RCF value **5796 x *g*** manually in the RCF field after selecting **RCF** in the same way.

4. Select **Time** by turning the knob. Press once, and then, by turning the knob again, set time as recommended in the particular protocol step. Confirm entry by pressing the knob.
5. For the Centrifuge 4 16KS, set the temperature to 40°C.
6. Open the lid, place the 96-well plates with the metal carriers in the buckets, and then close the lid. The start and lid keys light up.
7. Push **Start** to start the centrifuge. When the centrifuge is running, the lid key will not be lit. Each run can be interrupted by pushing **Stop**.
8. At the end of the run, the lid key will light up. Open the centrifuge lid by pressing the lid key. Remove the plates. All preset parameters remain after a run has finished.

Lysate filtration with QIAshredder (DNeasy Mini and Maxi procedures)

In the DNeasy Plant Mini and Maxi procedures, cell debris and salt precipitates are removed by centrifugation through a QIAshredder spin column. The preparation of a cleared lysate is essential to prevent clogging of the DNeasy spin column in the following step. Traditional methods involve removing the debris and precipitates by centrifugation and using the supernatant in subsequent steps. However, not all particulate matter forms a compact pellet; this makes preparation of a cleared lysate by centrifugation very difficult. The QIAshredder spin column removes all cell debris and precipitates, thus making the preparation of a cleared lysate rapid and efficient.

With some starting materials (e.g., oak leaves), centrifugation of the entire lysate through the QIAshredder spin column can result in sheared DNA. Investigation has shown that this is not due to the pore size of the QIAshredder spin column, but rather due to the high viscosity of the lysate and the large amount of precipitates. These form a compact layer on the QIAshredder spin column. Centrifugation of the lysate through this layer can result in size reduction of the DNA. Therefore, for certain plant tissues, an additional centrifugation step is recommended. This additional centrifugation is part of the standard DNeasy Plant Maxi procedure and is included in the DNeasy Plant Mini procedure as an optional step.

Elution

Purified DNA is eluted from the DNeasy spin column or the DNeasy 96 plate using either Buffer AE or water. Optimal results are obtained by eluting twice. The elution volume is typically 2 x 100 µL for the DNeasy Plant Mini Kit and the DNeasy 96 Plant Kit, and 2 x 750 µL for the DNeasy Plant Maxi Kit.

Higher concentrations of DNA

If higher concentrations of DNA are required in the eluate, reducing the elution volume to 2 x 50 µL (Mini or DNeasy 96) or 2 x 500 µL (Maxi) significantly increases concentration but reduces overall yield (Table 4, Table 5, and Table 6). If larger amounts (Mini >20 µg; Maxi >100 µg) of DNA are loaded onto DNeasy Mini or Maxi spin columns, eluting with 2 x 200 µL (Mini) or 2 x 1000 µL (Maxi) will increase yield (Table 4 and Table 5).

Table 4. DNeasy mini procedure: elution volumes and corresponding yields

Source (100 mg young leaves)	Elution volume (µL)	Total DNA yield (µg)*	Concentration (ng/µL)
Arabidopsis	2 x 50	3.6	38
	2 x 100	3.8	20
	2 x 200	4.1	11
Barley	2 x 50	7.9	83
	2 x 100	9.5	50
	2 x 200	10.0	26
Tobacco	2 x 50	20.5	216
	2 x 100	23.2	122
	2 x 200	29.7	78

* Because the column has a certain residual volume, the volume of eluate recovered is always less than the volume of buffer used for elution; the actual yield is therefore less than the theoretical yield.

Table 5. DNeasy maxi procedure: elution volumes and corresponding yields

Source (1 g young leaves)	Elution volume (µL)	Total DNA yield (µg)*	Concentration (ng/µL)
Maize	2 x 500	92	102
	2 x 750	112	79
	2 x 1000	143	72
Fir	2 x 500	63	72
	2 x 750	90	65
	2 x 1000	93	48
Rape	2 x 500	21	23
	2 x 750	26	18
	2 x 1000	27	14

* Because the column has a certain residual volume, the volume of eluate recovered is always less than the volume of buffer used for elution; the actual yield is therefore less than the theoretical yield.

Table 6. DNeasy 96 procedure: elution volumes and corresponding yields

Source (1 g young leaves)	Elution volume (µL)	Total DNA yield (µg)*	Concentration (ng/µL)
Wheat	2 x 500	16.6	166
	2 x 100	20.4	102
Lupin	2 x 50	1.5	15
	2 x 100	2.0	10

* Because the plate has a certain residual volume, the volume of eluate recovered is always less than the volume of buffer used for elution; the actual yield is therefore less than the theoretical yield.

Preventing dilution of the eluate

The first eluate can contain up to 70% of the total DNA yield. Therefore, to prevent dilution of the first eluate, the second elution can be performed separately. To elute separately, the second eluate should be collected in a separate tube.

Composition of elution buffer

Buffer AE is 10 mM Tris-Cl, 0.5 mM EDTA, pH 9.0. The pH of Buffer AE is optimal for DNA elution from the DNeasy membrane. Elution with buffer of pH lower than 9.0 may reduce DNA

yield. For long-term storage of DNA, we recommend eluting in Buffer AE, because DNA stored in water is subject to acid hydrolysis.

DNA storage

DNA is stable for several weeks when stored at 2–8°C in Buffer AE. For long-term storage, freezing at –30°C to –15°C is recommended.

Protocol: Purification of Total DNA from Plant Tissue (Mini Protocol)

Important points before starting

- If using the DNeasy Plant Mini Kit for the first time, read “Important Notes” (page 18).
- Ensure that you are familiar with operating the TissueRuptor II or the TissueLyser. See “Disruption and homogenization using the TissueRuptor II” or “Disruption and homogenization using the TissueLyser system”. Refer to the *TissueRuptor II User Manual* or the *TissueLyser Handbook* for operating instructions.
- Buffer AP1 may develop a yellow color upon storage. This does not affect the procedure.
- All centrifugation steps are carried out at room temperature in a microcentrifuge.

Things to do before starting

- Buffer AP1 and Buffer AW1 concentrate may form precipitates upon storage. If necessary, warm to 65°C to redissolve (before adding ethanol to Buffer AW1). Do not heat Buffer AW1 after ethanol has been added.
- Buffer AW2 and Buffer AW1 are supplied as concentrates. Before using for the first time, add the appropriate amount of ethanol (96–100%) as indicated on the bottle to obtain a working solution.
- Preheat a water bath or heating block to 65°C.

Procedure

1. For disruption using the TissueRuptor II, follow step 2; for disruption using the TissueLyser, follow steps 3–7.

Alternatively, a plant or fungal tissue can be grounded to a fine powder under liquid nitrogen using a mortar and pestle. Transfer the tissue powder and liquid nitrogen to an appropriately sized tube and allow the liquid nitrogen to evaporate. Do not allow the sample to thaw. Proceed immediately to step 7.

2. **TissueRuptor II procedure:** Place the sample material (≤ 100 mg wet weight or ≤ 20 mg lyophilized tissue) into a 2 mL microcentrifuge tube. Add liquid nitrogen to the tube, and freeze the sample for 30 s. Keep the sample submerged in liquid nitrogen, and disrupt for approximately 30 s at full speed. Allow the liquid nitrogen to evaporate, and proceed immediately to step 7.

Alternatively, fresh or lyophilized material can be directly disrupted in lysis buffer (after step 7) without using liquid nitrogen, but this may cause shearing of high-molecular-weight DNA. We do not recommend disrupting frozen material in lysis buffer, because this can result in low yields and degraded DNA.

3. **TissueLyser procedure:** Place the sample material (≤ 100 mg wet weight or ≤ 20 mg lyophilized tissue) into a 2 mL safe-lock microcentrifuge tube, together with a 3 mm tungsten carbide bead. Freeze the tubes on dry ice for 30 s.

When using lyophilized tissue, the tubes do not need to be frozen on dry ice.

4. Place the tubes into the TissueLyser Adapter Set 2 x 24, and fix into the clamps of the TissueLyser. Immediately grind the samples for 1 min at 30 Hz.
5. Disassemble the adaptor set, remove the tubes, and refreeze on dry ice for 30 s.
When using lyophilized tissue, the tubes do not need to be frozen on dry ice.
6. Repeat step 4, reversing the position of the tubes within the adaptor set. Proceed immediately to step 7.

Note: To prevent variation in sample homogenization, the adaptor sets should be removed from the TissueLyser and disassembled after the first disruption step. For the

second disruption step, the adaptor sets should be reassembled so that the tube order is reversed. Rotating the racks of tubes in this way ensures that all samples are thoroughly and equally disrupted.

Note: The majority of plant tissue is ground to a fine powder after two disruption steps. However, for some materials, one disruption step may be sufficient. Other tissues, such as seeds and roots, may require three disruption steps. Optimization of the disruption procedure may be required for some plant material.

7. Add 400 μL Buffer AP1 and 4 μL RNase A stock solution (100 mg/mL) to a maximum of 100 mg (wet weight) or 20 mg (dried) disrupted plant or fungal tissue and vortex vigorously.

No tissue clumps should be visible. Vortex or pipette further to remove any clumps. Clumps of tissue will not lyse properly and will therefore result in a lower yield of DNA. In rare cases, where clumps cannot be removed by pipetting and vortexing, a disposable micropestle may be used.

Note: Do not mix Buffer AP1 and RNase A before use.

8. Incubate the mixture for 10 min at 65°C. Mix 2–3 times during incubation by inverting tube. This step lyses the cells.
9. Add 130 μL Buffer P3 to the lysate. Mix and incubate for 5 min on ice.

This step precipitates detergent, proteins, and polysaccharides.

10. **Recommended:** Centrifuge the lysate for 5 min at 20,000 $\times g$ (14,000 rpm).

Some plant materials can generate very viscous lysates and large amounts of precipitates during this step. This can result in shearing of the DNA in the next step (see “Lysate filtration with QIAshredder (DNeasy Mini and Maxi procedures)”, page 25). In this case, optimal results are obtained if the majority of these precipitates are removed by centrifugation for 5 min at 20,000 $\times g$ (14,000 rpm). After centrifugation, apply supernatant to QIAshredder Mini spin column and continue with step 11.

11. Pipette the lysate into the QIAshredder Mini spin column (lilac) placed in a 2 mL collection tube, and centrifuge for 2 min at 20,000 $\times g$ (14,000 rpm).

It may be necessary to cut off the end of the pipette tip to apply the lysate to the QIAshredder Mini spin column. The QIAshredder Mini spin column removes most precipitates and cell debris, but a small amount will pass through and form a pellet in the collection tube. Be careful not to disturb this pellet in step 12.

12. Transfer the flow-through fraction from step 11 into a new tube (not supplied) without disturbing the cell-debris pellet.

Typically 450 μL lysate is recovered. For some plant species less lysate is recovered. In this case, determine the volume for the next step.

13. Add 1.5 volumes of Buffer AW1 to the cleared lysate, and mix by pipetting.

For example, to 450 μL lysate, add 675 μL Buffer AW1. Reduce the amount of Buffer AW1 accordingly if the volume of lysate is smaller. A precipitate may form after the addition of Buffer AW1, but this will not affect the DNeasy procedure.

Note: Ensure that ethanol has been added to Buffer AW1.

Note: It is important to pipette Buffer AW1 directly onto the cleared lysate and to mix immediately.

14. Pipette 650 μL of the mixture from step 13, including any precipitate that may have formed, into the DNeasy Mini spin column placed in a 2 mL collection tube (supplied). Centrifuge for 1 min at $\geq 6000 \times g$ (corresponds to ≥ 8000 rpm for most microcentrifuges), and discard the flow-through. * Reuse the collection tube in step 15.

15. Repeat step 14 with remaining sample. Discard flow-through* and collection tube.

16. Place the DNeasy Mini spin column into a new 2 mL collection tube (supplied), add 500 μL Buffer AW2 and centrifuge for 1 min at $\geq 6000 \times g$ (≥ 8000 rpm). Discard the flow-through and reuse the collection tube in step 17.

Note: Ensure that ethanol is added to Buffer AW2.

17. Add 500 μL Buffer AW2 to the DNeasy Mini spin column, and centrifuge for 2 min at $20,000 \times g$ (14,000 rpm) to dry the membrane.

* Flow-through fractions contain Buffer AW1 and are therefore not compatible with bleach. See "Safety Information".

It is important to dry the membrane of the DNeasy Mini spin column, because residual ethanol may interfere with subsequent reactions. This centrifugation step ensures that no residual ethanol will be carried over during elution. Discard flow-through and collection tube.

After washing with Buffer AW2, the DNeasy Mini spin column membrane is usually only slightly colored. In the rare case that the membrane remains significantly colored after washing with Buffer AW2, refer to “Darkly colored membrane or green/yellow eluate after washing with Buffer AW2” in the Troubleshooting Guide on page 53.

Note: After centrifugation, remove the DNeasy Mini spin column from the collection tube carefully so that the column does not come into contact with the flow-through, because that will result in carryover of ethanol.

18. Transfer the DNeasy Mini spin column to a 1.5 mL or 2 mL microcentrifuge tube (not supplied), and pipette 100 μ L Buffer AE directly onto the DNeasy membrane. Incubate for 5 min at room temperature, and then centrifuge for 1 min at $\geq 6000 \times g$ (≥ 8000 rpm) to elute.

Elution with 50 μ L (instead of 100 μ L) increases the final DNA concentration in the eluate significantly but also reduces overall DNA yield. If larger amounts of DNA (>20 μ g) are loaded, eluting with 200 μ L (instead of 100 μ L) increases yield. See “Elution”, page 26.

19. Repeat step 18 once.

A new microcentrifuge tube can be used for the second elution step to prevent dilution of the first eluate. Alternatively, the microcentrifuge tube can be reused for the second elution step to combine the eluates. See “Elution”, page 26.

Note: More than 200 μ L should not be eluted into a 1.5 mL microcentrifuge tube because the DNeasy Mini spin column will come into contact with the eluate.

Protocol: Purification of Total DNA from Plant Tissue (Maxi Protocol)

Important points before starting

- If using the DNeasy Plant Maxi Kit for the first time, read “Important Notes” (page 18).
- Ensure that you are familiar with operating the TissueRuptor II or the TissueLyser. See “19” or “20”. Refer to the *TissueRuptor II User Manual* or the *TissueLyser Handbook* for operating instructions.
- Buffer AP1 may develop a yellow color upon storage. This does not affect the procedure.
- All centrifugation steps are carried out at 3000–5000 $\times g$ (although 5000 $\times g$ is preferable) at room temperature in a laboratory centrifuge with swing-out rotor. Do not use a fixed-angle rotor; see “Equipment and Reagents to be Supplied by User”.

Things to do before starting

- Buffer AW1 concentrate may form precipitates upon storage. If necessary, warm to 65°C to redissolve (before adding ethanol). Do not heat Buffer AW1 after ethanol has been added.
- Preheat Buffer AP1 to 65°C. This heating is necessary for the DNeasy Plant Maxi procedure and will also dissolve any precipitate that may have formed in Buffer AP1.
- Buffer AW2 and Buffer AW1 are supplied as concentrates. Before using for the first time, add the appropriate amount of ethanol (96–100%) as indicated on the bottle to obtain a working solution.
- Preheat a water bath or a heating block to 65°C.

Procedure

1. For disruption using the TissueRuptor II, follow step 2; for disruption using the TissueLyser, follow steps 3–7.

Alternatively, plant or fungal tissue can be ground to a fine powder under liquid nitrogen using a mortar and pestle. Transfer the tissue powder and liquid nitrogen to an appropriately sized tube and allow the liquid nitrogen to evaporate. Do not allow the sample to thaw. Proceed immediately to step 8.

2. **TissueRuptor II procedure:** Place the sample material (≤ 1 g wet weight or ≤ 0.2 g lyophilized tissue) into a 15 mL centrifuge tube. Add liquid nitrogen to the tube and freeze the sample for 30 s. Keep the sample submerged in liquid nitrogen, and disrupt at full speed until the sample is homogenized. Allow the liquid nitrogen to evaporate, and proceed immediately to step 8.

Disruption time depends on the starting material used. Keep the disruption time as short as possible to avoid shearing of genomic DNA.

Alternatively, fresh or lyophilized material can be directly disrupted in lysis buffer (after step 8) without using liquid nitrogen, but this may cause shearing of high-molecular-weight DNA. We do not recommend disrupting frozen material in lysis buffer, because this can result in low yields and degraded DNA.

3. **TissueLyser procedure:** Place the sample material (≤ 1 g wet weight or ≤ 0.2 g lyophilized tissue) into a grinding jar, together with the stainless steel grinding ball.

Freeze the grinding jar in liquid nitrogen for approximately 30 s. When using lyophilized tissue, the grinding jars do not need to be frozen in liquid nitrogen.

4. Place the grinding jars into the clamps of the TissueLyser. Immediately grind the samples for 1 min at 30 Hz.
5. Remove the grinding jars, and refreeze in liquid nitrogen for 30 s. When using lyophilized tissue, the samples do not need to be frozen in liquid nitrogen.
6. Repeat step 4.
7. Transfer the fine powder into a 15 mL centrifuge tube (not supplied) using a spatula.

Do not allow the sample to thaw. Proceed immediately to step 8.

Note: The majority of plant tissue is ground to a fine powder after 2 disruption steps. However, for some materials 1 disruption step may be sufficient. Other tissues (such as seeds and roots) may require 3 disruption steps. Optimization of the disruption procedure may be required for some plant material.

8. Add 5 mL Buffer AP1 (preheated to 65°C) and 10 µL RNase A stock solution (100 mg/mL) to a maximum of 1 g (wet weight) or 0.2 g (dried) disrupted plant or fungal tissue and vortex vigorously.

No tissue clumps should be visible. Vortex or pipette further to remove any clumps. Clumps of tissue will not lyse properly and will therefore result in a lower yield of DNA. In rare cases where clumps cannot be removed by pipetting and vortexing, a disposable micropestle may be used.

Note: Do not mix Buffer AP1 and RNase A before use.

9. Incubate the mixture for 10 min at 65°C. Mix 2–3 times during incubation by inverting tube. This step lyses the cells.
10. Add 1.8 mL Buffer P3 to the lysate. Mix and incubate for 10 min on ice.
This step precipitates detergent, proteins, and polysaccharides.
11. Centrifuge the lysate at 3000–5000 $\times g$ for 5 min at room temperature.
A pellet will form, but some particles will float.
12. Decant the supernatant into a QIAshredder Maxi spin column (lilac) placed in a 50 mL collection tube, and centrifuge at 3000–5000 $\times g$ for 5 min at room temperature in a swing-out rotor. Transfer the flow-through in the collection tube, without disturbing the pellet, into a new 50 mL tube (not supplied). Record the volume.
Typically, 5–6 mL of lysate is recovered. After centrifugation of the sample, most of the debris and precipitates will be retained in the filter, but there will also be a pellet in the collection tube. Avoid disturbing the pellet when transferring the supernatant.
13. Add 1.5 volumes of Buffer AW1 to the cleared lysate, and mix immediately by vortexing.

For example, to 5 mL cleared lysate, add 7.5 mL Buffer AW1. Reduce the amount of Buffer AW1 accordingly if the volume of lysate is smaller. A precipitate may form after the addition of Buffer AW1, but this will not affect the DNeasy procedure.

Note: Ensure that ethanol has been added to Buffer AW1.

Note: It is important to pipette Buffer AW1 directly onto the cleared lysate and mix immediately.

14. Pipette the sample (maximum 15 mL), including any precipitate that may have formed, into the DNeasy Maxi spin column placed in a 50 mL collection tube (supplied). Centrifuge at 3000–5000 $\times g$ for 5 min at room temperature in a swing-out rotor. Discard the flow-through. *

Reuse the collection tube in step 15.

15. Add 12 mL Buffer AW2 to the DNeasy Maxi spin column, and centrifuge for 10 min at 3000–5000 $\times g$ to dry the membrane. Discard flow-through and collection tube.

Note: Ensure that ethanol has been added to Buffer AW2.

It is important to dry the membrane of the DNeasy Maxi spin column, because residual ethanol may interfere with subsequent reactions. This centrifugation step ensures that no residual ethanol will be carried over during elution.

After washing with Buffer AW2, the DNeasy Maxi spin column membrane is usually only slightly colored. In the rare case that the membrane remains significantly colored after washing with Buffer AW2, refer to “Darkly colored membrane or green/yellow eluate after washing with Buffer AW2” in the “Troubleshooting Guide” on page 53.

16. Transfer the DNeasy Maxi spin column to a new 50 mL tube (supplied). Pipette 0.75–1 mL Buffer AE directly onto the DNeasy Maxi spin column membrane. Incubate for 5 min at room temperature, and then centrifuge for 5 min at 3000–5000 $\times g$ to elute.

Note: Elution may also be performed with 0.5 mL of Buffer AE (instead of 0.75–1 mL). This increases the final DNA concentration in the eluate, but also reduces overall DNA yield. See “Elution”, page 26.

17. Add another 0.75–1 mL of Buffer AE and repeat the elution step as described in step 16. The first and second eluates may be combined or collected separately. For separate collection of the eluates, see “Elution”, page 26.

* Flow-through fractions contain Buffer AW1 and are therefore not compatible with bleach. See “Safety Information”.

Protocol: Purification of Total DNA from Fresh Plant Tissue (DNeasy 96 Protocol)

Important points before starting

- If using the DNeasy Plant 96 Kit for the first time, read “Important Notes” (page 18).
- Ensure that you are familiar with operating the TissueLyser and the QIAGEN 96-Well-Plate Centrifugation System. See “Disruption and homogenization using the TissueLyser system” and “Centrifugation (DNeasy 96 procedures)”. Refer to the *TissueLyser Handbook* and the centrifuge user manual for operating instructions.
- This protocol describes processing of 192 samples (2 x 96). If you wish to process 96 or fewer samples, provide a balance for the TissueLyser by assembling a second plate sandwich using a rack of collection microtubes without samples or buffers, but containing tungsten carbide beads, and fixing this second sandwich into the empty clamp.
- Tungsten carbide beads are reusable. See “Appendix B: Recovery and Cleaning of Beads and S-Blocks” for recovery and cleaning details.
- All centrifugation steps should be performed at room temperature. If the Centrifuge 4 16KS is used, set the temperature to 40°C for all centrifugation steps.
- DNA can appear as a smear on agarose gels when using this protocol. This can be avoided by using “Protocol: Purification of Total DNA from Fresh Plant Tissue (DNeasy 96 Protocol)”.

Things to do before starting

- Buffer AW2 and Buffer AW1 are supplied as concentrates. Before using for the first time, add the appropriate amount of ethanol (96–100%) as indicated on the bottle to obtain a working solution.

- Buffer AW1 concentrate may form precipitates upon storage. If necessary, warm to 65°C to redissolve (before adding ethanol). Do not heat Buffer AW1 after ethanol has been added.
- Preheat Buffer AP1 to 65°C. This heating is necessary for the DNeasy 96 Plant procedure and will also dissolve any precipitate that may have formed in Buffer AP1.

Procedure

1. Harvest leaves and place up to 50 mg into every tube in the 2 collection microtube racks.

Unless a different optimal amount of starting material has been previously determined, do not use more than 50 mg (wet weight) per sample (see “Sample size”, page 18). Most leaf material can be stored at 2–8°C for at least 24 h prior to processing without affecting DNA yield or quality.

Keep the clear covers from the collection microtube racks for use in step 10.

Use the plate register cards provided to record the position of each sample in the racks.

2. Add one tungsten carbide bead to each collection microtube.
3. Combine Buffer AP1, RNase A, and Reagent DX to make a working lysis solution (Table [7](#)). Pipette 400 µL working lysis solution into each collection microtube. Seal the microtubes with the caps provided.

It is important to prepare a fresh working lysis solution. To allow thorough mixing of the solution, combine the components in a tube and vortex to mix. Then, dispense the solution into a reagent reservoir for use with a multichannel pipette.

Table 7. Preparation of a working lysis solution

Reagent	Volume per sample	Volume for 2 x 96 samples*
Buffer AP1 (preheated to 65°C)	400 µL	90 mL
RNase A (100 mg/mL)	1 µL	225 µL
Reagent DX†	1 µL	225 µL

* 15% excess mixture is included in these calculations to allow for pipetting errors.

† Reagent DX is viscous.

- Sandwich each rack of collection microtubes between adapter plates and fix into TissueLyser clamps as described in the TissueLyser user manual.

Note: Ensure that the microtubes are properly sealed with caps.

Important: Two plate sandwiches must be clamped to the TissueLyser to provide balance. To process 96 samples or fewer, assemble a second plate sandwich using a rack of collection microtubes that contain tungsten carbide beads but no samples or buffers, and fix it into the empty clamp.

- Grind the samples for 1.5 min at 30 Hz.

Important: Prolonging the disruption time may result in DNA shearing.

- Remove and disassemble the plate sandwiches. Ensure that the collection microtubes are tightly closed. Reassemble the plate sandwiches so that the collection microtubes nearest the TissueLyser in steps 4 and 5 are now outermost. Reinsert the plate sandwiches into the TissueLyser.

Rotating the racks of collection microtubes in this way ensures that all samples are thoroughly disrupted (see “Disruption and homogenization using the TissueLyser system”). More foam will form in the tubes that were outermost during the initial disruption step.

Important: Merely rotating the entire plate sandwich so that the QIAGEN logos are upside down when reinserted into the mixer mill is not sufficient, because the same samples that were outermost during the initial disruption will remain outermost in the second disruption step.

- Grind the samples for another 1.5 min at 30 Hz.

Important: Prolonging the disruption time may result in shearing of DNA.

8. Remove the plate sandwiches from the TissueLyser and remove the adapter plates from each rack of collection microtubes. To collect any solution from the caps, centrifuge the collection microtubes. Allow the centrifuge to reach 3000 rpm, and then stop the centrifuge.

Do not prolong this step.

9. Remove and discard caps. Add 130 μ L Buffer P3 to each collection microtube.
10. Close the microtubes carefully with new caps (provided); ensure that the microtubes are properly sealed to avoid leakage during shaking. Place a clear cover (saved from step 1) over each rack of collection microtubes and shake the racks vigorously up and down for 15 s. To collect any solution from the caps, centrifuge the collection microtubes. Allow the centrifuge to reach 3000 rpm and then stop the centrifuge.

Do not prolong this step.

Note: To ensure optimal DNA yields, it is important to shake the racks of collection microtubes vigorously up and down with both hands for the full 15 s. The genomic DNA will not be sheared by vigorous shaking.

The centrifugation step prevents precipitates from freezing to the caps, which would otherwise be difficult to remove after incubation at -20°C (step 11).

Keep the clear covers from the collection microtube racks for use in step 15.

11. Incubate the racks of collection microtubes for 10 min at -20°C .

This incubation aids the precipitation of proteins and inhibitors of downstream applications following addition of Buffer P3.

12. Centrifuge the racks of collection microtubes for 5 min at 6000 rpm.

Compact pellets will form, but some particles may float. Be careful not to transfer any of these particles in the following step.

13. Remove and discard the caps. Carefully transfer 400 μ L of each supernatant to new racks of collection microtubes (provided), ensuring that the new tubes are in the correct orientation.

Do not discard the pellets, because they contain the tungsten carbide beads, which can be recovered and reused (see “Appendix B: Recovery and Cleaning of Beads and S-Blocks”).

Do not transfer more than 400 μL of the supernatant, because otherwise the capacity of the DNeasy 96 plates and the S-Blocks used in subsequent steps will be exceeded.

If less than 400 μL supernatant is recovered, adjust the amount of Buffer AW1 in step 14 accordingly.

Collection microtubes are connected in strips of 8. To avoid transferring particulate matter, it is helpful to remove the strips from the rack so that the contents of the microtubes are visible, and to use a multichannel pipette on its lowest speed setting. Save the used collection microtubes to recover the tungsten carbide beads at a later stage (see “Appendix B: Recovery and Cleaning of Beads and S-Blocks”).

14. Add 1.5 volumes (typically 600 μL) Buffer AW1 to each sample.

Note: Ensure that ethanol has been added to Buffer AW1.

A white precipitate may form upon addition of Buffer AW1. This precipitate does not interfere with the DNeasy 96 Plant procedure or any subsequent application.

15. Close the collection microtubes with new caps (provided); ensure that the tubes are properly sealed to prevent leakage during shaking. Place a clear cover over each rack of collection microtubes and shake the racks vigorously up and down for 15 s.

To collect any solution from the caps, centrifuge the collection microtubes. Allow the centrifuge to reach 3000 rpm, and then stop the centrifuge.

Do not prolong this step.

Note: To ensure optimal DNA yields, it is important to shake the racks of collection microtubes vigorously up and down with both hands for the full 15 s. The genomic DNA will not be sheared by vigorous shaking.

16. Place 2 DNeasy 96 plates on top of S-Blocks (provided). Mark the DNeasy 96 plates for later sample identification.

17. Remove and discard the caps from the collection microtubes. Carefully transfer 1 mL of each sample to each well of the DNeasy 96 plates.

Take care not to wet the rims of the wells to avoid aerosols during centrifugation.

Do not transfer more than 1 mL per well.

Note: Lowering the pipette tips to the bottoms of the wells may cause sample overflow and cross-contamination. Therefore, remove one set of caps at a time, and begin drawing up the samples as soon as the pipette tips come in contact with the liquid. Repeat until all the samples have been transferred to the DNeasy 96 plates.

18. Seal each DNeasy 96 plate with an AirPore Tape Sheet (provided). Centrifuge for 4 min at 6000 rpm.

AirPore Tape prevents cross-contamination between samples during centrifugation.

After centrifugation, check that all of the lysate has passed through the membrane in each well of the DNeasy 96 plates. If lysate remains in any of the wells, centrifuge for a further 4 min.

19. Remove the tape. Carefully add 800 μ L Buffer AW2 to each sample.

Note: Ensure that ethanol has been added to Buffer AW2.

20. Centrifuge for 15 min at 6000 rpm to dry the DNeasy membranes.

For efficient drying, do not reseal the DNeasy 96 plate with AirPore Tape.

Important: Residual ethanol in the DNeasy membranes derived from Buffer AW2 may inhibit PCR and must be removed by centrifugation before elution of the DNA.

Note: DNeasy membranes are sometimes slightly colored after this wash step. This should not affect the DNeasy 96 Plant procedure. A very dark membrane may indicate that too much starting material was used. A second wash step with 800 μ L ethanol (96–100%) may improve DNA quality in these cases. Empty the flow-through from the S-Block before performing this second wash step.

21. To elute the DNA, place each DNeasy 96 plate in the correct orientation on a new rack of Elution Microtubes RS (provided), add 100 μ L Buffer AE to each sample and seal the

DNeasy 96 plates with new AirPore Tape Sheets (provided). Incubate for 1 min at room temperature. Centrifuge for 2 min at 6000 rpm.

Elution in 2 x 50 μL (instead of 2 x 100 μL) increases DNA concentration, but decreases the overall DNA yield (see "Elution", page 26).

22. Repeat step 21 with another 100 μL Buffer AE.

Use new caps (provided) to seal the Elution Microtubes RS for storage.

Protocol: Purification of Total DNA from Frozen or Lyophilized Plant Tissue (DNeasy 96 Protocol)

Important points before starting

- If using the DNeasy 96 Plant Kit for the first time, read “Important Notes” (page 18).
- Ensure that you are familiar with operating the TissueLyser and the QIAGEN 96-Well-Plate Centrifugation System. See “Disruption and homogenization using the TissueLyser system” and “Centrifugation (DNeasy 96 procedures)”. Refer to the *TissueLyser Handbook* and the centrifuge user manual for operating instructions.
- Do not allow the TissueLyser adapter plates to come into contact with liquid nitrogen during the procedure.
- This protocol describes processing of 192 samples (2 x 96). If you wish to process 96 or fewer samples, provide a balance for the TissueLyser by assembling a second plate sandwich using a rack of collection microtubes without samples or buffers, but containing tungsten carbide beads, and fixing this second sandwich into the empty clamp.
- Tungsten carbide beads are reusable. See “Appendix B: Recovery and Cleaning of Beads and S-Blocks” for recovery and cleaning details.
- All centrifugation steps should be performed at room temperature. If the Centrifuge 4 16KS is used, set the temperature to 40°C for all centrifugation steps.

Things to do before starting

- Buffer AW2 and Buffer AW1 are supplied as concentrates. Before using for the first time, add the appropriate amount of ethanol (96–100%) as indicated on the bottle to obtain a working solution.
- Buffer AW1 concentrate may form precipitates upon storage. If necessary, warm to 65°C to redissolve (before adding ethanol). Do not heat Buffer AW1 after ethanol has been added.

- Preheat Buffer AP1 to 80°C. This heating is necessary for the DNeasy 96 Plant procedure, and will also dissolve any precipitate that may have formed in Buffer AP1. (When using lyophilized tissue without using liquid nitrogen, preheat Buffer AP1 to 65°C.)
- If possible, harvest plant samples directly into the collection microtubes. Freeze or lyophilize the harvested samples before starting the protocol.

Procedure

1. Place sample material (≤ 50 mg wet weight or ≤ 10 mg lyophilized tissue) into every tube in the 2 collection microtube racks.

Unless a different optimal amount of starting material has been previously determined, do not use more than 50 mg (wet weight) or 10 mg (lyophilized tissue) per sample (see “Sample size”, page 18).

Important: Do not allow frozen sample material to thaw during handling and weighing. Keep the racks of collection microtubes on dry ice to keep samples from thawing.*

Keep the clear covers from the collection microtube racks, for use in step 4.

Use the plate register cards provided to record the position of each sample in the racks.

2. Add one tungsten carbide bead to each collection microtube. Seal the microtubes with the caps provided.
3. Cool the racks of collection microtubes in on dry ice. Ensure that the microtubes remain tightly closed.

When using lyophilized tissue, the microtubes do not need to be frozen on dry ice. Continue with step 4.

4. Place a clear cover (saved from step 1) over each rack of collection microtubes, and knock the racks upside down against the bench 5 times to ensure that all tungsten carbide beads can move freely within the microtubes. But do not allow the leaf material to thaw. Remove the clear cover.

During freezing on dry ice, the leaf material and bead in each collection microtube may stick together, hindering disruption in the TissueLyser. This step ensures that the beads are free for optimal disruption.

Keep the clear covers from the collection microtube racks, for use in step 8.

5. Sandwich each rack of collection microtubes between adapter plates and fix into TissueLyser clamps as described in the TissueLyser user manual. Work quickly so that the plant material does not thaw.

Note: Ensure that the microtubes are properly sealed with caps.

Important: Do not allow the TissueLyser adapter plates to come into contact with liquid nitrogen.

Important: Two plate sandwiches must be clamped to the TissueLyser to provide balance. To process 96 samples or fewer, assemble a second plate sandwich using a rack of collection microtubes that contain tungsten carbide beads but no samples or buffers, and fix it into the empty clamp.

6. Grind the samples for 1 min at 20 Hz.

Important: Prolonging the disruption time may result in DNA shearing.

7. Remove and disassemble the plate sandwiches, noting the orientation of the racks of collection microtubes during the first round of disruption. Ensure that the collection microtubes are tightly closed.

8. Cool the racks of collection microtubes again on dry ice. Place a clear cover over each rack of collection microtubes and knock the racks upside down against the bench 5 times to ensure that all tungsten carbide beads can move freely within the microtubes. But do not allow the leaf material to thaw. Remove the clear cover.

Important: Do not put the adapter plates on dry ice. Disassemble the plate sandwiches as described above, and place only the racks of collection microtubes on dry ice. When using lyophilized tissue, the microtubes do not need to be frozen on dry ice. Continue with step 9.

Keep the clear covers from the collection microtube racks, for use in step 13.

9. Ensure that the collection microtubes are tightly closed. Reassemble the plate sandwiches so that the collection microtubes nearest the TissueLyser in steps 5 and 6 are now outermost. Reinsert the plate sandwiches into the TissueLyser. Work quickly so that the plant material does not thaw.

Rotating the racks of collection microtubes in this way ensures that all samples are thoroughly disrupted (See “Disruption and homogenization using the TissueLyser system”).

Important: Merely rotating the entire plate sandwich so that the QIAGEN logos are upside down when reinserted into the mixer mill is not sufficient, because the same samples that were outermost during the initial disruption will remain outermost in the second disruption step.

10. Grind the samples for another 1 min at 20 Hz.
11. Remove the plate sandwiches from the TissueLyser and remove the adapter plates from each rack of collection microtubes. Knock the racks against the bench 5 times to ensure that no tissue powder remains in the caps. Keep the samples frozen until working lysis solution is added (step 12).

Important: The samples should not be allowed to thaw while the working lysis solution is being prepared (step 12). Store the samples at -20°C until the working lysis solution is ready.

12. Combine Buffer AP1, RNase A, and Reagent DX according to the scheme below, to make a working lysis solution (Table 8). Carefully remove the caps from the collection microtubes. Immediately pipette 400 μL working lysis solution into each collection microtube.

It is important to prepare a fresh working lysis solution. To allow thorough mixing of the solution, combine the components in a tube and vortex to mix. Then, dispense the solution into a reagent reservoir for use with a multichannel pipette.

If the working lysis solution is not used immediately and is allowed to cool, precipitates may form when the solution is added to the disrupted leaf material. If this occurs, redissolve the precipitates by incubating the racks of collection microtubes in a water

bath at 65°C for 10–20 min. Place a heavy plate over the racks during incubation to prevent the caps from coming off. Remove all water that has entered the racks before centrifuging in the following step.

Prior to opening the microtubes, knock the racks against a bench to remove any tissue powder from the caps (step 11). The microtubes may become brittle at low temperature.

Take care not to break the connection between the tubes when removing the caps.

Table 8. Preparation of a working lysis solution

Reagent	Volume per sample	Volume for 2 x 96 samples*
Buffer AP1 (preheated to 80°C) [†]	400 µL	90 mL
RNase A (100 mg/mL)	1 µL	225 µL
Reagent DX [‡]	1 µL	225 µL

* 15% excess mixture is included in these calculations to allow for pipetting errors.

[†] When using lyophilized tissue without using liquid nitrogen, preheat Buffer AP1 to 65°C.

[‡] Reagent DX is viscous.

13. Seal the microtubes with new caps (provided); ensure that the microtubes are properly sealed to avoid leakage during shaking. Place a clear cover over each rack of collection microtubes, and shake the racks vigorously up and down for 15 s. To collect any solution from the caps, centrifuge the collection microtubes. Allow the centrifuge to reach 3000 rpm, and then stop the centrifuge.

Do not prolong this step.

Note: To ensure optimal DNA yields, it is important to shake the racks of collection microtubes vigorously up and down with both hands for the full 15 s. The genomic DNA will not be sheared by vigorous shaking.

Keep the clear covers from the collection microtube racks, for use in step 15.

14. Remove and discard caps. Add 130 µL Buffer P3 to each collection microtube.
15. Close the microtubes carefully with new caps (provided); ensure that the microtubes are properly sealed to avoid leakage during shaking. Place a clear cover over each rack of collection microtubes, and shake the racks vigorously up and down for 15 s. To collect

any solution from the caps, centrifuge the collection microtubes. Allow the centrifuge to reach 3000 rpm, and then stop the centrifuge.

Do not prolong this step.

Note: To ensure optimal DNA yields, it is important to shake the racks of collection microtubes vigorously up and down with both hands for the full 15 s. The genomic DNA will not be sheared by vigorous shaking.

The centrifugation step prevents precipitates from freezing to the caps, which would otherwise be difficult to remove after incubation at -20°C (step 16).

Keep the clear covers from the collection microtube racks for use in step 20.

16. Incubate the racks of collection microtubes for 10 min at -20°C .

This incubation aids the precipitation of proteins and inhibitors of downstream applications following addition of Buffer P3.

17. Centrifuge the racks of collection microtubes for 5 min at 6000 rpm.

Compact pellets will form, but some particles may float. Be careful not to transfer any of these particles in the following step.

18. Remove and discard the caps. Carefully transfer 400 μL of each supernatant to new racks of collection microtubes (provided), ensuring that the new tubes are in the correct orientation.

Do not discard the pellets, because they contain the tungsten carbide beads, which can be recovered and reused (see "Appendix B: Recovery and Cleaning of Beads and S-Blocks").

Do not transfer more than 400 μL of the supernatant, because otherwise the capacity of the DNeasy 96 plates and the S-Blocks used in subsequent steps will be exceeded.

If less than 400 μL supernatant is recovered, adjust the amount of Buffer AW1 in step 19 accordingly.

Collection microtubes are connected in strips of 8. To avoid transferring particulate matter, it is helpful to remove the strips from the rack, so that the contents of the microtubes are visible, and to use a multichannel pipette on its lowest speed setting.

Save the used collection microtubes to recover the tungsten carbide beads at a later stage (see “Appendix B: Recovery and Cleaning of Beads and S-Blocks”).

19. Add 1.5 volumes (typically 600 μ L) of Buffer AW1 to each sample.

Note: Ensure that ethanol has been added to Buffer AW1.

A white precipitate may form upon addition of Buffer AW1. This precipitate does not interfere with the DNeasy 96 Plant procedure or any subsequent application.

20. Close the collection microtubes with new caps (provided); ensure that the tubes are properly sealed to prevent leakage during shaking. Place a clear cover over each rack of collection microtubes, and then shake the racks vigorously up and down for 15 s. To collect any solution from the caps, centrifuge the collection microtubes. Allow the centrifuge to reach 3000 rpm, and then stop the centrifuge.

Do not prolong this step.

Note: To ensure optimal DNA yields, it is important to shake the racks of collection microtubes vigorously up and down with both hands for the full 15 s. The genomic DNA will not be sheared by vigorous shaking.

21. Place two DNeasy 96 plates on top of S-Blocks (provided). Mark the DNeasy 96 plates for later sample identification.
22. Remove and discard the caps from the collection microtubes. Carefully transfer 1 mL of each sample to each well of the DNeasy 96 plates.

Take care not to wet the rims of the wells to avoid aerosols during centrifugation. Do not transfer more than 1 mL per well.

Note: Lowering pipette tips to the bottoms of the wells may cause sample overflow and cross-contamination. Therefore, remove one set of caps at a time, and begin drawing up the samples as soon as the pipette tips contact the liquid. Repeat until all the samples have been transferred to the DNeasy 96 plates.

23. Seal each DNeasy 96 plate with an AirPore Tape Sheet (provided). Centrifuge for 4 min at 6000 rpm.

AirPore Tape prevents cross-contamination between samples during centrifugation.

After centrifugation, check that all of the lysate has passed through the membrane in each well of the DNeasy 96 plates. If lysate remains in any of the wells, centrifuge for a further 4 min.

24. Remove the tape. Carefully add 800 μ L Buffer AW2 to each sample.

Note: Ensure that ethanol has been added to Buffer AW2 prior to use.

25. Centrifuge for 15 min at 6000 rpm to dry the DNeasy membranes.

For efficient drying, do not reseal the DNeasy 96 plate with AirPore Tape.

Important: Residual ethanol in the DNeasy membranes derived from Buffer AW2 may inhibit PCR and must be removed by centrifugation before elution of the DNA.

Note: DNeasy membranes are sometimes slightly colored after this wash step. This should not affect the DNeasy 96 Plant procedure. A very dark membrane may indicate that too much starting material was used. A second wash step with 800 μ L ethanol (96–100%) may improve DNA quality in these cases. Empty the flow-through from the S-Block before performing this second wash step.

26. To elute the DNA, place each DNeasy 96 plate in the correct orientation on a new rack of Elution Microtubes RS (provided), add 100 μ L Buffer AE to each sample, and seal the DNeasy 96 plates with new AirPore Tape Sheets (provided). Incubate for 1 min at room temperature. Centrifuge for 2 min at 6000 rpm.

Elution in 2 x 50 μ L (instead of 2 x 100 μ L) increases DNA concentration but decreases the overall DNA yield (see “Elution”, page 26).

27. Repeat step 26 with another 100 μ L Buffer AE.

Use new caps (provided) to seal the Elution Microtubes RS for storage.

Troubleshooting Guide

This troubleshooting guide may be helpful in solving any problems that may arise. For more information, see also the Frequently Asked Questions page at our Technical Support Center: www.qiagen.com/FAQ/FAQList.aspx. The scientists in QIAGEN Technical Services are always happy to answer any questions you may have about either the information and protocols in this handbook or sample and assay technologies (for contact information, visit www.qiagen.com).

Comments and suggestions

Clogged QIAshredder spin column

- | | | |
|----|---|---|
| a) | Insufficient centrifugation | Increase the g -force and centrifugation time. |
| b) | Mini protocol: High viscosity of lysate, precipitates | Perform the optional centrifugation step before loading a large amount of the lysate onto the QIAshredder, as outlined in step 10 of "Protocol: Purification of Total DNA from Plant Tissue (Mini Protocol)" (page 29). |

Clogged DNeasy membrane

- | | | |
|----|-----------------------------------|--|
| a) | Carryover of particulate material | In future preparations, ensure that no particulate material is transferred following centrifugation through the QIAshredder spin column (Mini and Maxi procedures) or when supernatants are transferred to new microtubes prior to addition of Buffer AW1 (DNeasy 96 procedures) |
| b) | Lysate too viscous | Reduce the amount of starting material and/or increase the amounts of Buffer AP1 and Buffer P3. |
| c) | Insufficient centrifugation | Increase the g -force and centrifugation time. |

Low yield

- | | | |
|----|------------------------------|--|
| a) | Insufficient disruption | Ensure that the starting material is completely disrupted. See "Disruption and homogenization using the TissueRuptor II" and "Disruption and homogenization using the TissueLyser system". |
| b) | Insufficient lysis | Reduce the amount of starting material and/or increase the amounts of Buffer AP1 and Buffer P3. |
| c) | Incorrect binding conditions | Make sure that the amount of lysate is accurately determined so that the correct amount of Buffer AW1 is added to adjust binding conditions correctly. |

Comments and suggestions

- | | | |
|----|---|--|
| d) | DNA still bound to the membrane | Increase the volume of Buffer AE or water to 200 μ L (DNeasy Plant Mini and DNeasy 96 procedures) or to \geq 1 mL (DNeasy Plant Maxi procedure), and incubate on the column for 5 min at room temperature before centrifugation. |
| e) | Maxi protocol: Incorrect centrifugation method | Use a swinging-bucket rotor. Do not use a fixed-angle rotor. |

DNA sheared

- | | | |
|----|--|--|
| a) | Precipitate has formed in Buffer AP1 | Ensure that any precipitate that has formed in Buffer AP1 is completely dissolved before use, by heating to 65°C if necessary. |
| b) | Mini protocol: Debris and precipitates in lysate | Perform the optional centrifugation step before loading a large amount of the lysate onto the QIAshredder spin column, as described in step 10 of "Protocol: Purification of Total DNA from Plant Tissue (Mini Protocol)". |

Darkly colored membrane or green/yellow eluate after washing with Buffer AW2

- | | | |
|----|--|--|
| a) | Too much starting material | Reduce the amount of starting material in future preps. |
| b) | Mini protocol: Insufficient washing of the membrane | After washing with Buffer AW2 (step 17), perform an additional wash with 500 μ L ethanol (96–100%). Centrifuge for 2 min at 20,000 $\times g$ (14,000 rpm) to dry the membrane. Continue with step 18 of "Protocol: Purification of Total DNA from Plant Tissue (Mini Protocol)". |
| c) | Maxi protocol: Insufficient washing of the membrane | After washing with Buffer AW2 (step 15), perform an additional wash with 12 mL ethanol (90% v/v in water). Centrifuge for 10 min at 3000–5000 $\times g$ and dry the column for 15–30 min at 65°C in an oven to remove residual ethanol. Continue with step 16 of "Protocol: Purification of Total DNA from Plant Tissue (Maxi Protocol)". |

DNA does not perform well in downstream experiments

- | | | |
|----|--|---|
| a) | Ethanol carryover | Ensure that during the second wash with Buffer AW2, the DNeasy spin column or DNeasy 96 plate is centrifuged at 20,000 $\times g$ (14,000 rpm) for 2 min (Mini), at 3000–5000 $\times g$ for 10 min (Maxi), or at full speed for 15 min (DNeasy 96) to dry the membrane. After centrifugation, remove the DNeasy spin column or DNeasy 96 plate carefully from the collection tubes so that the column or plate does not come into contact with the flow-through, because this will result in carryover of ethanol. |
| b) | Salt carryover | Ensure that Buffer AW2 is at room temperature before use. |
| c) | Insufficient/excess DNA used in downstream application | Optimize the amount of DNA used in the downstream application, if necessary. Downstream applications can be adversely affected by insufficient or excess DNA. |

Comments and suggestions

- d) **DNeasy 96 protocols:** Eluate is slightly green or yellow
- In future preparations, include an additional slightly green or yellow wash with 800 μL ethanol (96–100%) after washing with Buffer AW2. If 2 wash steps are performed, the first centrifugation step should be 5 min at 5600 $\times g$, after which the flow-through should be discarded from the S-Block before performing the second wash step. Ensure that residual ethanol is removed during the second wash step by centrifuging for 15 min at maximum speed. Slight color in the eluate will not necessarily interfere with downstream applications.

DNeasy 96 protocols: Yields vary across DNeasy 96 plate

- a) Varying amount of starting material across the DNeasy 96 plate
- Adjust the amounts of starting materials accordingly.
- b) Racks of collection microtubes not turned during disruption (when using the TissueLyser)
- It is essential to turn the racks of collection microtubes during disruption in the TissueLyser to ensure all samples are evenly disrupted (see “Disruption and homogenization using the TissueLyser system”).
- c) Non-uniform sample disruption when using alternative disruption methods
- Ensure that all samples are uniformly disrupted.

Contact Information

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Appendix A: Determination of Yield, Purity, and Length of DNA

Determination of yield and purity

The concentration and purity of DNA can be determined by measuring the absorbance at 260 nm (A_{260}) and 280 nm (A_{280}) in a spectrophotometer. To ensure accuracy, make sure the absorbance readings fall into the linear range of your method (e.g., between 0.1 and 1.0 for spectrophotometric OD). Sample dilution should be adjusted accordingly. An absorbance of 1.0 at 260 nm corresponds to 50 μg of DNA per milliliter ($A_{260} = 1 = 50 \mu\text{g}/\text{mL}$). We recommend scanning absorbance from 220–320 nm, because this will indicate whether other factors are interfering with absorbance at 260 and 280 nm.

DNA samples from plant tissue often contain copurified polysaccharides and other metabolites, which can interfere with OD readings. Absorbance scans should show a symmetric peak at 260 nm and have an overall smooth shape (Figure 4). Purity is determined by calculating the ratio of absorbance at 260 nm to absorbance at 280 nm. Pure DNA has an A_{260}/A_{280} ratio of 1.7–1.9.

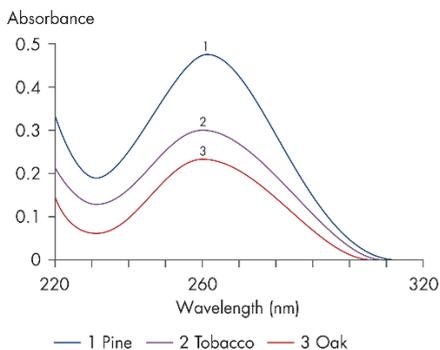


Figure 4. UV scan of DNeasy purified DNA diluted 1:5 in water.

If both DNA and RNA are present in the same sample, both will be measured with a spectrophotometer. If DNA alone is to be quantified in a sample that also contains RNA, a fluorimeter must be used. DNA purified using DNeasy Plant procedures is free of RNA contamination, because an RNase digestion step is included in the procedure.

Average DNA yields are shown in Table 9.

Table 9. Average DNA yields obtained with DNeasy Plant Kits

Source		Yield (µg DNA) per 100 mg*
Arabidopsis	<i>Arabidopsis thaliana</i>	3–4
Barley	<i>Hordeum vulgare</i>	8–12
Fir	<i>Abies alba</i>	6–10
Maize	<i>Zea mays</i>	15–20
Oak	<i>Quercus robur</i>	10–15
Pine	<i>Pinus sylvestris</i>	20–25
Potato	<i>Solanum tuberosum</i>	4–6
Rape	<i>Brassica napus</i>	2–4
Spinach	<i>Spinacia oleracea</i>	5–10
Tobacco	<i>Nicotiana tabacum</i>	20–25
Tomato	<i>Lycopersicon esculentum</i>	10–15

* DNA yields vary due to genome size, ploidy, age of sample, etc. All material was collected as young leaves or needles.

Determination of length

The precise length of genomic DNA should be determined by pulse-field gel electrophoresis (PFGE) through an agarose gel. To prepare the sample for PFGE, the DNA should be concentrated by alcohol precipitation and the DNA pellet dried briefly at room temperature for 5–10 minutes. Avoid drying the DNA pellet for more than 10 min, because over-dried genomic DNA is very difficult to redissolve. Redissolve in approximately 30 µL TE buffer, pH

8.0,3F for at least 30 min at 60°C. Load 3–5 µg DNA per well. Standard PFGE conditions are as follows:

- 1% agarose gel in 0.5x TBE electrophoresis buffer*
- Switch intervals = 5–40 s
- Run time = 17 h
- Voltage = 170 V

* When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate safety data sheets (SDSs), available from the product supplier.

Appendix B: Recovery and Cleaning of Beads and S-Blocks

Cleaning beads and grinding balls

Tungsten carbide and stainless steel beads and grinding balls can be reused. Used beads can be recovered from cell debris and cleaned using the procedure below:

1. Follow the appropriate step for DNeasy Mini, Maxi, or 96 below:

DNeasy Mini: Close the cap of the 2 mL collection tube, and briefly vortex to dislodge the bead and pellet from the bottom of the tube.

DNeasy Maxi: If using grinding jars for disruption of large sample volumes, skip to step 2.

DNeasy 96: Seal the collection microtubes with caps. Place a clear cover (saved from step 1 of DNeasy 96 Plant procedures) over each rack of collection microtubes, turn the racks upside down, and knock them against the bench 5 times to free the tungsten carbide beads from the surrounding material.

2. Empty the contents of tubes/jars into a sieve and rinse the beads thoroughly with water.
3. Incubate beads in 0.4 M HCl* for 1 min at room temperature to degrade any DNA and avoid cross-contamination in future preparations.
4. Rinse beads thoroughly with distilled water to remove the HCl.
5. Dry beads before use.

* When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate safety data sheets (SDSs), available from the product supplier.

Cleaning S-Blocks

To avoid cross-contamination, rinse the S-Blocks thoroughly in tap water after each use, incubate for 1 min at room temperature in 0.4 M HCl, empty, and wash thoroughly with distilled water. Used S-Blocks can also be autoclaved after washing. Additional S-Blocks can be ordered separately.

Ordering Information

Product	Contents	Cat. no.
DNeasy Plant Mini Kit (50)	50 DNeasy Mini Spin Columns, 50 QIAshredder Mini Spin Columns, RNase A, buffers, collection tubes (2 mL)	69104
DNeasy Plant Mini Kit (250)	250 DNeasy Mini Spin Columns, 250 QIAshredder Mini Spin Columns, RNase A, buffers, collection tubes (2 mL)	69106
DNeasy Plant Maxi Kit (24)	24 DNeasy Maxi Spin Columns, 24 QIAshredder Maxi Spin Columns, RNase A, buffers, collection tubes (50 mL)	68163
DNeasy 96 Plant Kit (6)	6 DNeasy 96 Plates, buffers, reagents, RNase A, S-Blocks, collection microtubes (1.2 mL), caps, AirPore tape sheets	69181
DNeasy Plant Pro Kit (50)	Tissue Disruption Tubes, MB Spin Columns, Buffers, Collection Tubes (1.5 and 2 mL), for 50 preps	69204
DNeasy Plant Pro Kit (250)	Tissue Disruption Tubes, MB Spin Columns, Buffers, Collection Tubes (1.5 and 2 mL), for 250 preps	69206
DNeasy Plant Mini Kit (50)	50 DNeasy Mini Spin Columns, 50 QIAshredder Mini Spin Columns, RNase A, buffers, collection tubes (2 mL)	69104
DNeasy Plant Mini Kit (250)	250 DNeasy Mini Spin Columns, 250 QIAshredder Mini Spin Columns, RNase A, buffers, collection tubes (2 mL)	69106

Product	Contents	Cat. no.
TissueRuptor II		
TissueRuptor II	Handheld rotor–stator homogenizer, 5 TissueRuptor Disposable Probes	Varies
TissueRuptor Disposable Probes (25)	25 nonsterile plastic disposable probes for use with the TissueRuptor II (and the old TissueRuptor)	990890
TissueLyser system		
TissueLyser III	Universal laboratory mixer-mill disruptor	Varies
TissueLyser LT	Compact bead mill, 100–240 V AC, 50–60 Hz; requires the TissueLyser LT Adapter, 12-Tube (available separately)	85600
TissueLyser LT Adapter, 12 Tube	Adapter for disruption of up to 12 samples in 2 mL microcentrifuge tubes on the TissueLyser LT	69980
TissueLyser Adapter Set 2 x 24	Two sets of adapter plates and 2 racks for use with 2 mL microcentrifuge tubes on the TissueLyser III & II	69982
TissueLyser Adapter Set 2 x 96	2 sets of adapter plates for use with collection microtubes (racked) on the TissueLyser III & II (Collection Microtubes [racked] not included)	69984
Grinding Jar Set, Stainless Steel (2 x 10 mL)	2 grinding jars (10 mL), 2 stainless steel grinding balls (20 mm)	69985
Tungsten Carbide Beads, 3 mm (200)	200 tungsten carbide beads (3 mm diameter), suitable for use with TissueLyser systems	69997
QIAGEN 96-Well Plate Centrifugation System		

Product	Contents	Cat. no.
Centrifuge 4 16S	Universal laboratory centrifuge with brushless motor	Varies
Centrifuge 4 16KS	Refrigerated universal laboratory centrifuge with brushless motor	Varies
Accessories		
Collection Tubes (2 mL)	1000 collection tubes (2 mL)	19201
Collection Microtubes (racked, 10 x 96)	Nonsterile polypropylene tubes (1.2 mL), 960 in racks of 96	19560
Collection Microtube Caps (120 x 8)	Nonsterile polypropylene caps for collection microtubes (1.2 mL) and round-well blocks, 960 in strips of 8	19566
S-Blocks (24)	96-well blocks with 2.2 mL wells, 24 per case	19585
AirPore Tape Sheets (50)	Microporous tape sheets for covering 96 well blocks: 50 sheets per pack	19571
Buffer AW2 (Concentrate, 324 mL)	324 mL wash buffer (2) concentrate	19072
Buffer AE (240 mL)	240 mL Elution Buffer for 1000 preps	19077
QIAcube Connect – for fully automated nucleic acid extraction with QIAGEN spin-column kits		
QIAcube Connect*	Instrument, connectivity package, 1 year warranty on parts and labor	Inquire
Starter Pack, QIAcube	Filter-tips, 200 µL (1024), 1000 µL filter-tips (1024), 30 mL reagent bottles (12), rotor adapters (240), 1.5 mL elution tubes (240), rotor adapter holder (1)	990395

Product	Contents	Cat. no.
Related Products		
BioSprint® 15 DNA Plant Kit (360)	For 360 preps: 5-rod covers, 5 Tube MagAttract® Suspension G, buffers, and reagents	941517
BioSprint 96 DNA Plant Kit (576)*	For 576 preps: Large 96-Rod Covers, 96-Well Microplates MP, S Blocks, MagAttract Suspension G, buffers and reagents	941557
MagAttract 96 DNA Plant Core Kit (24)	MagAttract suspension and buffers for 24 x 96 preps	67163
RNeasy® Plant Mini Kit (50)	For 50 RNA minipreps: 50 RNeasy Mini Spin Columns, 50 QIAshredder Mini Spin Columns, collection tubes (1.5 mL and 2 mL), and RNase-free reagents and buffers	74904

* All QIAcube Connect instruments are provided with a region-specific connectivity package, including tablet and equipment necessary to connect to the local network. Further, QIAGEN offers comprehensive instrument service products, including service agreements, installation, introductory training and preventive subscription. Contact your local sales representative to learn about your options.

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Document Revision History

Revision	Description
February 2020	Updated text, ordering information, and intended use for QIAcube Connect.
July 2020	Replaced liquid nitrogen with dry ice in steps 3 and 5 of "Protocol: Purification of Total DNA from Plant Tissue (Mini Protocol)", for consistency with the TissueLyser handbook.
March 2024	Updated the following: <ul style="list-style-type: none">• Updated cat.no. for TissueLyser III• Replaced liquid nitrogen with dry ice of "Protocol: Purification of Total DNA from Plant Tissue (DNeasy 96 Protocol)", for consistency with the TissueLyser handbook.

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