Quick-Start Protocol M

March 2016

AllPrep® DNA/RNA FFPE Kit, Part 1

The AllPrep DNA/RNA FFPE Kit (cat. no. 80234) can be stored for at least 9 months if not otherwise stated on label: buffers at room temperature (15–25°C); other components at 2–8°C.

Further information

- AllPrep DNA/RNA FFPE Handbook: www.qiagen.com/HB-0373
- Safety Data Sheets: www.qiagen.com/safety
- Technical assistance: support.qiagen.com

Notes before starting

- Unless otherwise stated, perform all steps quickly at room temperature (15–25°C).
- Unless otherwise stated, centrifugation is performed at ≥8000 x g (≥10,000 rpm).
- Reconstitute Buffer FRN, Buffer RPE, Buffer AW1, Buffer AW2 and RNase-Free DNase I as described in the handbook. Mix by shaking before use.
- Set a thermal mixer, heated orbital incubator, or heating block to 56°C.
- Flow-through from steps 8, 9 and 12 contains Buffer RLT and Buffer FRN and is therefore not compatible with bleach.
- Symbols: purification of total RNA that does not include small RNAs; purification of total RNA that does include small RNAs.
- 1. Prepare sections and remove paraffin as described in the handbook.
- 2. Resuspend the pellet obtained after deparaffinization by adding 150 µl Buffer PKD and flicking the tube to loosen the pellet. Add 10 µl proteinase K and mix by vortexing.
- 3. Incubate at 56°C for 15 min. Incubate on ice for 3 min.
- 4. Centrifuge for 15 min at 20,000 x g (14,000 rpm). Carefully transfer the supernatant, without disturbing the pellet, to a new 1.5 ml or ▲ 2 ml Safe-Lock microcentrifuge tube (see handbook for ordering information) for RNA purification (below).

5. Keep the pellet for DNA purification; see "Genomic DNA purification" in the *Quick-Start Protocol AllPrep DNA/RNA FFPE Kit, Part 2*.

RNA purification

- 6. Incubate the supernatant at 80°C for 15 min.
- 7. Add 320 µl Buffer RLT, and mix by vortexing or pipetting. Add 720 µl or ▲ 1120 µl ethanol (96–100%), and mix well by vortexing or pipetting.
- 8. Transfer 700 µl sample, including any precipitate, to an RNeasy® MinElute® spin column placed in a 2 ml collection tube (supplied). Centrifuge for 15 s.
 Repeat step until complete lysate is used.
- 9. Add 350 µl Buffer FRN to the spin column. Centrifuge for 15 s.
- 10.Add 10 μ l DNase I stock solution to 70 μ l Buffer RDD. Mix by gently inverting the tube. Add the DNase I incubation mix (80 μ l) directly to the spin column membrane, and place on the benchtop (20–30°C) for 15 min.
- 11.Add 500 µl Buffer FRN to the spin column. Centrifuge for 15 s. Save the flow-through for use in step 12.
- 12.Place the spin column in a new 2 ml collection tube (supplied). Apply the flow-through from step 11 to the spin column. Centrifuge for 15 s.
- 13.Add 500 μ l Buffer RPE to the spin column. Centrifuge for 15 s. Repeat step.
- 14. Place the spin column in a new 2 ml collection tube (supplied). Open the lid, and centrifuge at full speed for 5 min to dry spin column membrane.
- 15. Place the spin column in a new 1.5 ml collection tube (supplied). Add 14–30 µl RNase-free water directly to the spin column membrane. Close lid, and incubate for 1 min at room temperature (15–25°C). Centrifuge at full speed for 1 min to elute the RNA.



Scan QR code for handbook.

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