

Guidelines for Laboratory Verification of Performance of QIAstat-Dx[®] GI Panel 2 Mini B&V

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Introduction

This document provides a sample protocol for the verification of QIAstat-Dx GI Panel 2 Mini B&V (cat. no. 691424). The protocol provides positive and negative tests for the pathogens detected by QIAstat-Dx GI Panel 2 Mini B&V.

Each laboratory is responsible for defining their verification procedure and ensuring that they meet regulatory, state and federal guidelines.

Table 1. Overview of sample verification method

Sample code	
Organism controls per sample control mix	2–3
Number of sample control mixes	2
Replicates per sample control mix	4
QIAstat-Dx Cartridges required	8
Expected number of positive results	20
Expected number of negative results	20
Approximate days of testing	2
Number of operators	2

Materials and methods

The procedure described in Table 1 and below generates multiple positive and negative results for each of the sample control mixes tested. The sample protocol was developed using NATtrol[™] Gastrointestinal 2 Verification Panel available from ZeptoMetrix Corporation (Buffalo, NY; cat. no. NATGIP2-QIA).

If testing is performed using a QIAstat-Dx Analyzer 2.0 with additional Analytical Modules or the QIAstat-Dx Rise, the laboratory director may choose not to perform the verification protocol on each Analytical Module. If the complete verification protocol is not performed on each Analytical Module, we advise distributing test replicates evenly among the different Analytical Modules of the system.

Table 2. Materials needed for the sample verification method

Material	Catalog number	Quantity
QIAstat-Dx GI Panel 2 Mini B&V kit (6 tests)	691424	2
QIAstat-Dx Operational Module*	9002813	1
QIAstat-Dx Analytical Module(s)	9002814	1–4
NATtrol Gastrointestinal 2 Verification Panel†	NATGIP2-QIA	1
Sample tubes, 2 mL	VWR 525-0131 (or similar)	4
Transfer pipettes	VWR 13-711-43 (or similar)	12

* Or QIAstat-Dx Rise Base (cat. no. 9003163)

† NATtrol Gastrointestinal 2 Verification Panel includes 4 x 1.0 mL of negative control made of a simulated stool matrix (stool diluent) that serves as sample diluent in the sample verification method. The kit provides 22 organism controls (each tube contains a volume of 0.250 mL), while preparation of control samples only requires those specified in Table 3.

Performance verification materials

The materials listed in Table 2 are required to perform verification with the sample protocol.

Sample verification method

This sample verification method describes how to prepare sample control mixes by mixing together either 2 or 3 different organism controls using a combination of pathogens from ZeptoMetrix NATtrol Gastrointestinal 2 Verification Panel. Proposed mixing of organism controls is provided in Table 3. The method tests a total of 8 sample control mixes (2 sample control mixes tested in 4 replicates each). For each assay run, the method provides 2–3 positive results and, correspondingly, 2–3 negative

results for the 5 targets in total that are detected and differentiated by QIAstat-Dx GI Panel 2 Mini B&V.

Alteration of this protocol should take into account additional lab personnel and the number of instruments, based on individual laboratory needs.

Mix the organism controls to create control samples at the beginning of the sample verification method. The sample control mixes can be stored at refrigerated conditions (2–8°C) for up to 3 days.

Note: It is important to prepare only the number of sample control mixes that will be tested within 3 days of preparation. The number of samples prepared may be increased or decreased based on the laboratory's work schedule and number of QIAstat-Dx Analyzer instruments.

Protocol

Day 1

1. Prepare Sample Control Mix 1 and Sample Control Mix 2 (refer to Table 3). Perform a quick step of vortex and spin to each ZeptoMetrix tube before use:
 - a. Transfer the appropriate volume of simulated stool matrix (provided as 4 x 1.0 mL negative control in the kit) to a new 2 mL tube.
 - b. Transfer 0.2 mL of each of the organism controls in the mix to the 2 mL tube.
 - c. Ensure the pooled sample is effectively mixed by vortexing prior to testing.
2. Test two replicates from Sample Control Mix 1. The duplicate samples should be tested in a single day by different operators (see Table 4).

3. Repeat step 2 for Sample Control Mix 2 to be tested on the same day.
4. Store the samples at 2–8°C for up to 3 days for the evaluation of day-to-day variation.

Day 2

To evaluate day-to-day variation, test the remaining volume of the sample control mixes prepared on Day 1 (Sample Control Mix 1 and Sample Control Mix 2) by repeating steps 2 and 3 above.

Table 3. Proposed organism-control mixing and expected results (positive or negative)

Mix	Organism	Organism control volume (mL)	Simulated stool matrix* volume (mL)	Final volume of mix (mL)	Expected results	Number of expected negative results
Sample Control Mix 1	<i>Salmonella enterica typhimurium</i>	0.2	0.9	1.5	Positive	2
	<i>Campylobacter jejuni</i>	0.2			Positive	
	<i>Shigella sonnei</i>	0.2			Positive	
Sample Control Mix 2	Norovirus GII	0.2	1.1	1.5	Positive	3
	Shiga-like toxin <i>Escherichia coli</i> EDL933	0.2			Positive	

* NATrol Gastrointestinal 2 Verification Panel includes 4 x 1.0 mL of negative control made of a simulated stool matrix (stool diluent) that serves as sample diluent in the sample verification method.

Table 4. Workflow for the sample verification method

	Day 1	Day 2
Operator 1	Sample Control Mix 1	Sample Control Mix 1
	Sample Control Mix 2	Sample Control Mix 2
Operator 2	Sample Control Mix 1	Sample Control Mix 1
	Sample Control Mix 2	Sample Control Mix 2

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