

Quick-Start Protocol

QIASprint® Pathogen Application

For use with QIASprint Connect

The QIASprint Pathogen PrepSet (cat. no. 585799) and the QIASprint Essential Kit A (384) (cat. no. 585009) are shipped at room temperature (15–25°C). All kit components should be stored dry at room temperature.

Further information

- QIASprint Pathogen Purification Guide: www.qiagen.com/HB-3728
- Safety Data Sheets: www.qiagen.com/safety
- Technical assistance: support.qiagen.com

Notes before starting

- Buffer ACL and Buffer AW1 contain guanidine and salt and are therefore not compatible with disinfecting reagents containing bleach.
- Prepare Buffer AW1 and Buffer AW2 according to the instructions on the bottle.
- Dissolve the lyophilized carrier RNA in RNase-free water to a final concentration of 1 µg/µL. See the *QIASprint Pathogen Purification Guide* (www.qiagen.com/HB-3728) for carrier RNA storage details.
- Ethanol (96–100%) is required in this protocol and needs to be supplied by the user.
- Isopropanol is required in this protocol and needs to be supplied by the user.
- For a 50 µL elution, add 75 µL RNase-free water; for 100 µL elution, add 125 µL. Select the appropriate protocol.
- If necessary, pretreat samples according to the appropriate procedure, as described in the *QIASprint Pathogen Purification Guide*.

Procedure

Sample preparation in sample plate

1. Prepare the lysis/binding master mix ($\times + 10\%$) according to the table below:

Lysis/binding mix	Per sample (µL)
Buffer ACL	400
Isopropanol	200
Carrier RNA	1
MagG Bead Suspension	25

Note: Ensure sufficient resuspension of the MagG Bead Suspension before distribution by thorough vortexing.

2. Add 20 µL Proteinase K into the bottom of a well of a prep plate insert (Sample Plate).
Important: Even though you can add QIAGEN Proteinase K after dispensing samples, it is best to add it beforehand.
Note: Do not add QIAGEN Proteinase K directly to Buffer ACL.
3. Transfer 200 µL of sample to the Sample Plate.
4. Add 626 µL of the lysis/binding mix to each well.
Note: Keep MagG Bead Suspension in suspension to ensure even distribution.

Plates preparation

5. Prepare the plates as described in the table below:

Plate	Content	Volume (µL)
Sample Plate	Proteinase K	20
	Sample	200
	Lysis/binding mix	626
Wash Plate 1	Buffer AW1	700
Wash Plate 2	Buffer AW2	700
Wash Plate 3	96–100% Ethanol	750
Elution Plate	RNase-free Water	75/125

Procedure on QIASprint Connect

6. Turn on the QIASprint Connect instrument and log in.
7. Navigate to the Protocols screen with the tab in the upper part of the home screen.
8. Locate the desired protocol for the Pathogen application, and press  and  **Run protocol** .
For additional information on protocol selection, please refer to the handbook.
Optional: Enter the run information in the Run details screen. This information will be displayed in the run report.
9. Close the hood and press **Confirm** to get to the loading screen.
10. With the plate hotels located on the lab bench, insert the plates according to the screen instructions into the hotels.
 - The prep cover must be located in the metal prep cover frame.
 - All prep plates need to sit firmly in their metal frames.
 - The QR codes need to face the user.
11. Open the hood and place the hotels on their platforms, taking care not to mix up the left and right hotels.
12. **Optional:** If you have heating or cooling steps during your protocol run and the thermal adapter is not already located on the heater-cooler, place the thermal adapter in one of the slots of the thermal adapter platform with the QR code facing the user. The instrument will install it automatically during the run.
13. Press **Execute run** to start the run.
14. After the end of the run, remove the plate hotels from the instrument. Store the elution plate safely or use it directly in your downstream applications.

Document Revision History

Date	Description
03/2026	Initial release
	Scan the QR code for handbook. For up-to-date licensing information and product-specific disclaimers, see the respective QIAGEN kit handbook or user manual.

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