

# GeneRead™ DNA FFPE Kit, Part 2

Uracil-N-glycosylase is shipped on dry ice and should be stored immediately upon receipt at  $-30$  to  $-15^{\circ}\text{C}$  in a constant-temperature freezer. When the product is stored under these conditions and handled correctly, performance is guaranteed until the expiration date (see the quality control label on Uracil-N-glycosylase tubes). Store QIAamp® MinElute® Columns at  $2$ – $8^{\circ}\text{C}$ . All other components of the GeneRead DNA FFPE Kit (cat. no. 180134) should be stored dry at room temperature ( $15$ – $25^{\circ}\text{C}$ ). Under these conditions, they are stable for at least 12 months if not otherwise stated on label.

## Further information

- *GeneRead DNA FFPE Handbook*: [www.qiagen.com/HB-1757](http://www.qiagen.com/HB-1757)
- Safety Data Sheets: [www.qiagen.com/safety](http://www.qiagen.com/safety)
- Technical assistance: [support.qiagen.com](http://support.qiagen.com)

## Notes before starting

- Unless stated otherwise, all centrifugation steps should be performed for 1 min at full speed (maximum  $20,000 \times g$ ) in a conventional, table-top centrifuge.
  - If Buffer AL contains precipitates, dissolve by heating to  $70^{\circ}\text{C}$  with gentle agitation.
  - Ethanol (96–100%) is not provided with the kit.
  - Add 25 ml ethanol (96–100%) to the bottle containing 19 ml Buffer AW1 concentrate.
  - Add 30 ml ethanol (96–100%) to the bottle containing 13 ml Buffer AW2 concentrate.
11. Add 250  $\mu\text{l}$  Buffer AL to the sample and mix thoroughly by vortexing. Then add 250  $\mu\text{l}$  ethanol (96–100%) to each sample and mix thoroughly by vortexing. Centrifuge briefly to remove drops from inside the lid.
12. Transfer 700  $\mu\text{l}$  lysate to the QIAamp MinElute column (in a 2 ml collection tube), close the lid and centrifuge. Discard the flow-through and reuse the collection tube.

13. Repeat step 12 until the complete lysate is used up.
14. Add 500  $\mu$ l Buffer AW1 to each spin column and centrifuge. Discard the flow-through and reuse the collection tube.
15. Add 500  $\mu$ l Buffer AW2 to each spin column and centrifuge. Discard the flow-through and reuse the collection tube.
16. Add 250  $\mu$ l ethanol (96–100%) to the spin column and centrifuge.
17. Discard the flow-through and collection tube.
18. Place the spin column into a new 2 ml collection tube (supplied) and centrifuge to remove any residual liquid.
19. Place the QIAamp MinElute column into a clean 1.5 ml microcentrifuge tube (not provided) and discard the collection tube containing the flow-through.
20. Open the lid of the QIAamp MinElute column and apply 20–40  $\mu$ l Buffer ATE to the center of the membrane.  
**IMPORTANT:** Ensure that Buffer ATE is equilibrated to room temperature. Dispense Buffer ATE onto the center of the membrane to ensure complete elution of bound DNA. The volume of eluate will be up to 5  $\mu$ l less than the volume of Buffer ATE that was applied to the column.
21. Close the lid and incubate at room temperature (15–25°C) for 1 min, then centrifuge.  
Incubating the QIAamp MinElute column loaded with Buffer ATE for 5 min at room temperature before centrifugation generally increases DNA yield.



Scan QR code for handbook.

For up-to-date licensing information and product-specific disclaimers, see the respective QIAGEN kit handbook or user manual.

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