

QIASymphony[®] RGQ Protocol Sheet

Settings to run the *artus*[®] CT/NG QS-RGQ Kit (Rotor-Gene[®] Q software 2.1)



Check availability of new electronic labeling revisions at www.qiagen.com/products/artusctngqsrgqkitce.aspx before test execution. The current revision status is indicated by the issue date (format: month/year).

General information



artus CT/NG QS-RGQ Kit

Version 1, REF 4569365

Important points before starting

- Take time to familiarize yourself with the Rotor-Gene Q before starting the protocol. See the instrument user manual.
- See also the *artus CT/NG QS-RGQ Kit Handbook* and relevant Application Sheet at www.qiagen.com/products/artusctngqsrgqkitce.aspx.
- Make sure that at the positive/negative controls as well as at least one negative control (NTC, provided in the kit and pipetted by the AS module) are included per PCR run. Because the *artus* CT/NG QS-RGQ Kit is a qualitative assay, no quantitation standards are supplied.

Procedure

1. **Place the PCR tubes in the 72-Well Rotor of the Rotor-Gene Q.**
2. **Place empty PCR tubes with lids in empty rotor positions.**
This will guarantee an optimal temperature distribution in the Rotor-Gene Q.
3. **Lock the rotor with the locking ring.**
4. **Transfer the cycler file from the QIASymphony AS to the Rotor-Gene Q computer.**

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5. Open the "New Run Wizard" dialog box (Figure 1). Check the "Locking Ring Attached" box and click "Next".

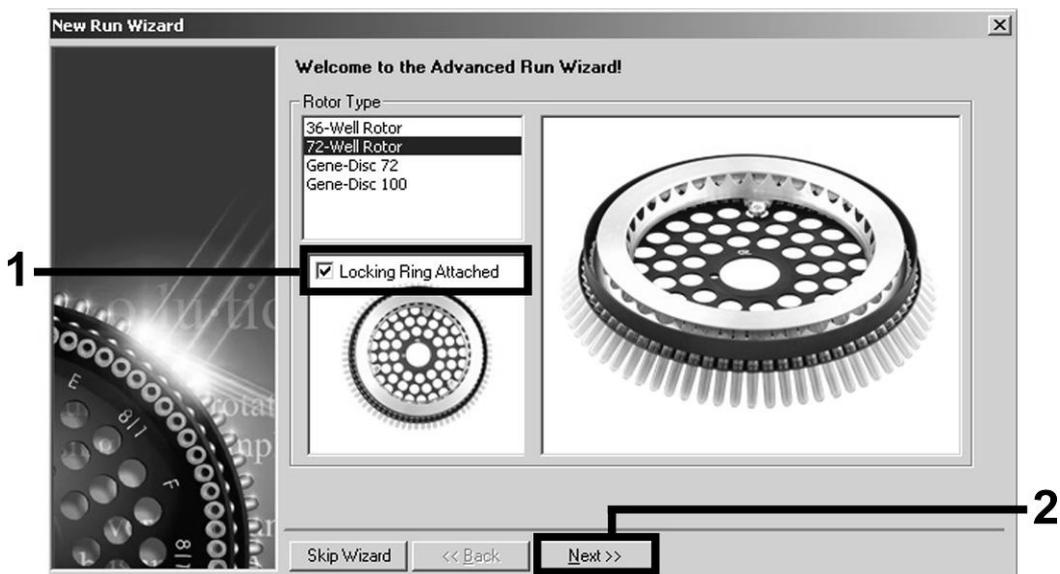


Figure 1. The "New Run Wizard" dialog box.

6. Select 25 for the PCR reaction volume and click "Next" (Figure 2).

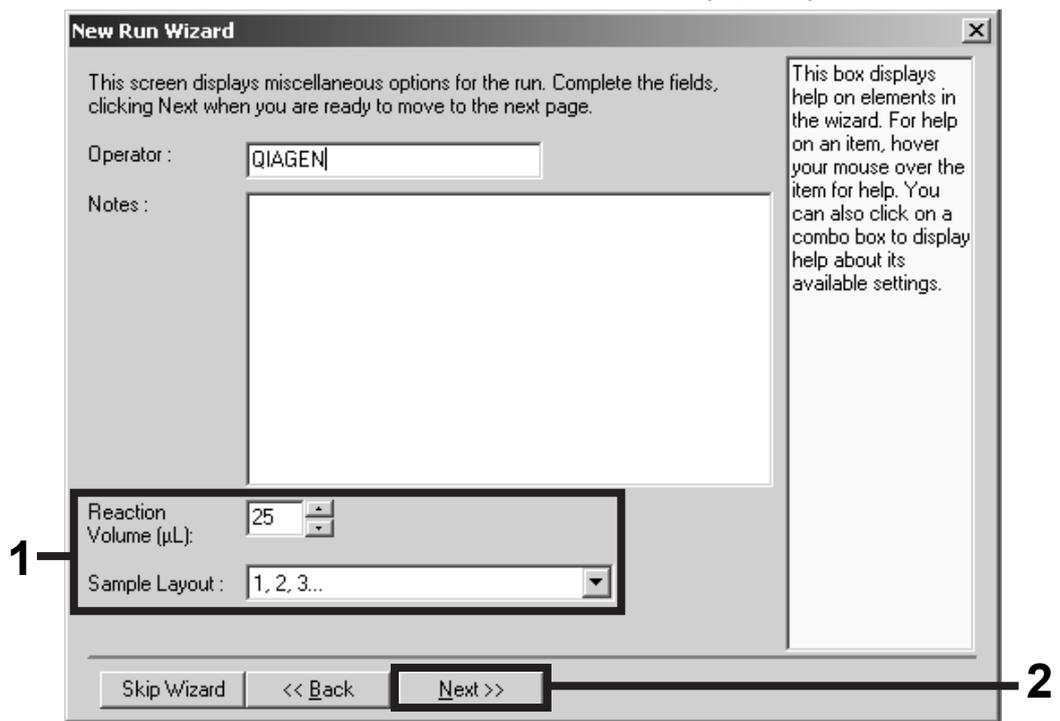


Figure 2. Setting the general assay parameters.

7. Click the "Edit Profile" button in the next "New Run Wizard" dialog box (Figure 3). Program the temperature profile as shown in Table 1, using the screenshots shown in Figures 3–5 as a guide.

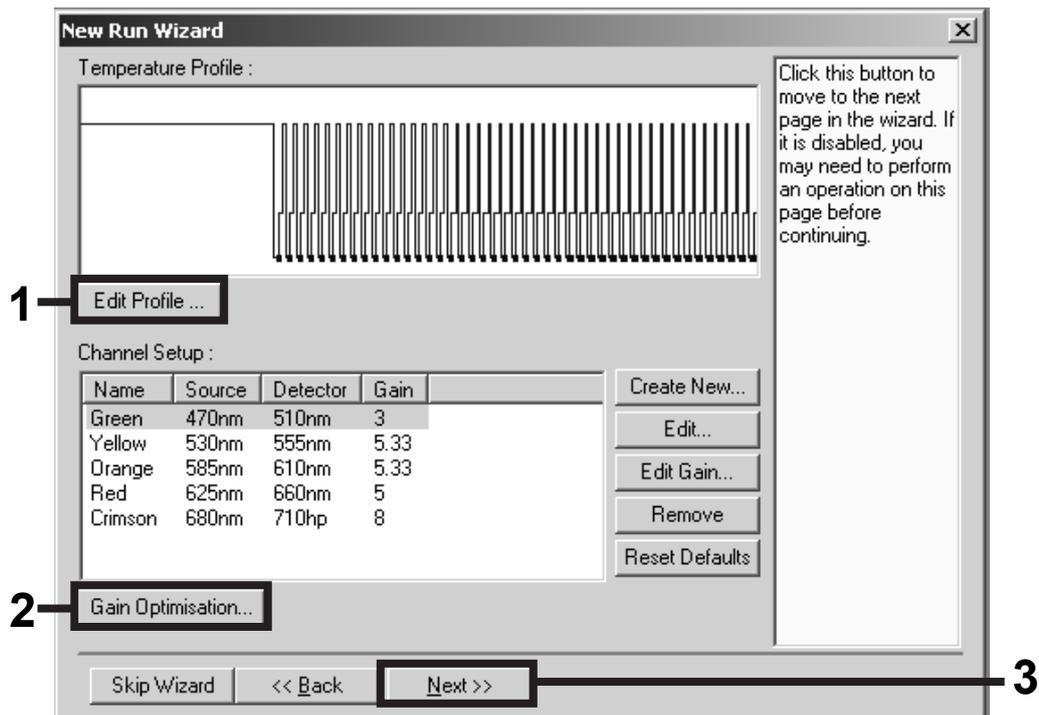


Figure 3. Editing the profile.

Table 1. Temperature profiles for the *artus* CT/NG QS-RGQ Kit

Hold	Temperature: 95 deg Time: 15 mins
Hold 2	Step not required
Cycling	45 times 95 deg for 11 secs 60 deg for 20 secs 72 deg for 20 secs

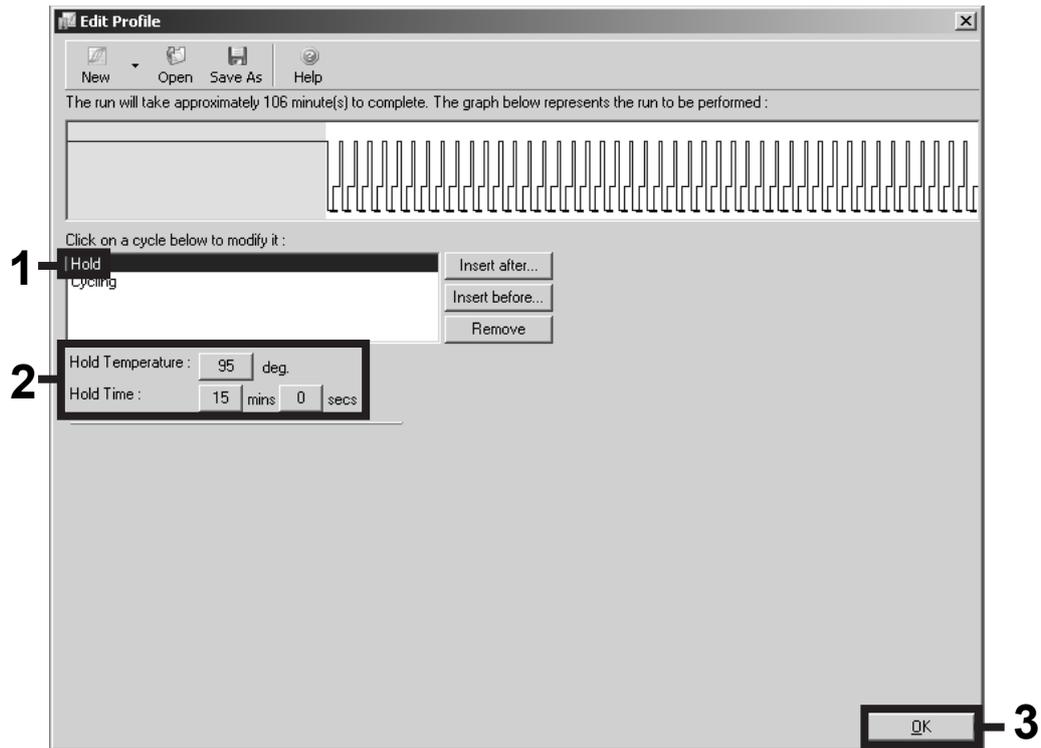


Figure 4. Initial activation of the hot-start enzyme.

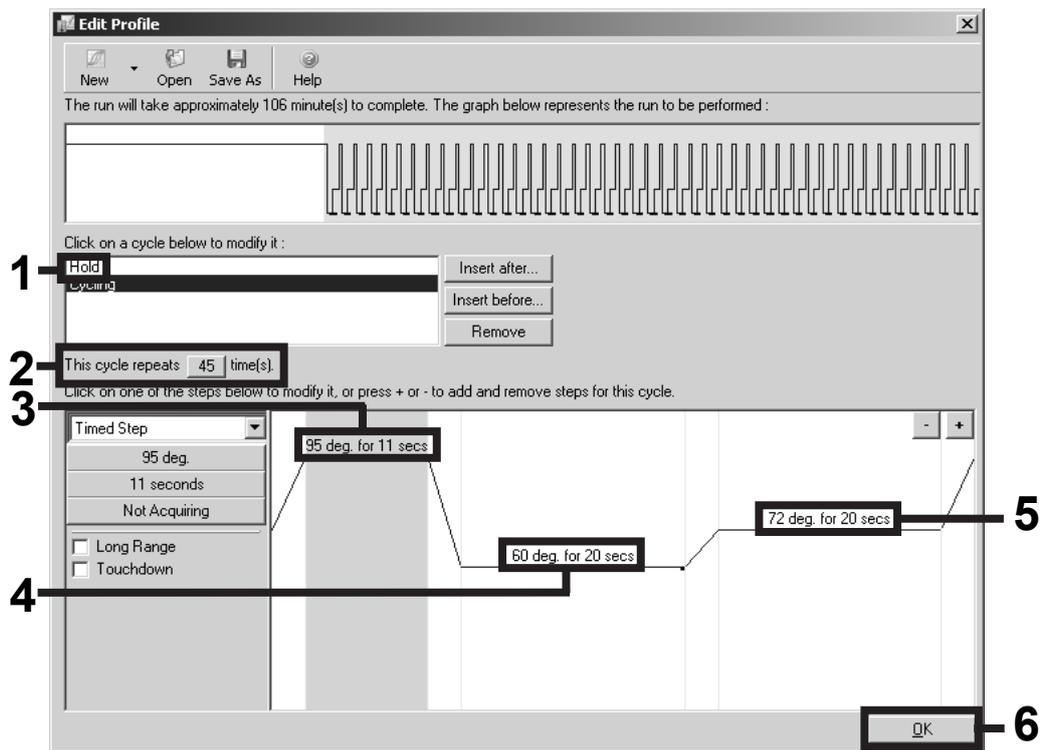


Figure 5. Amplification of the DNA.

- The detection range of the fluorescence channels has to be determined according to the fluorescence intensities in the PCR tubes. Click "Gain Optimisation" in the "New Run Wizard" dialog box (see Figure 3, page 3) to open the "Auto-Gain Optimisation Setup" dialog box. Set the calibration temperature to 60 to match the annealing temperature of the amplification program (Figure 6).

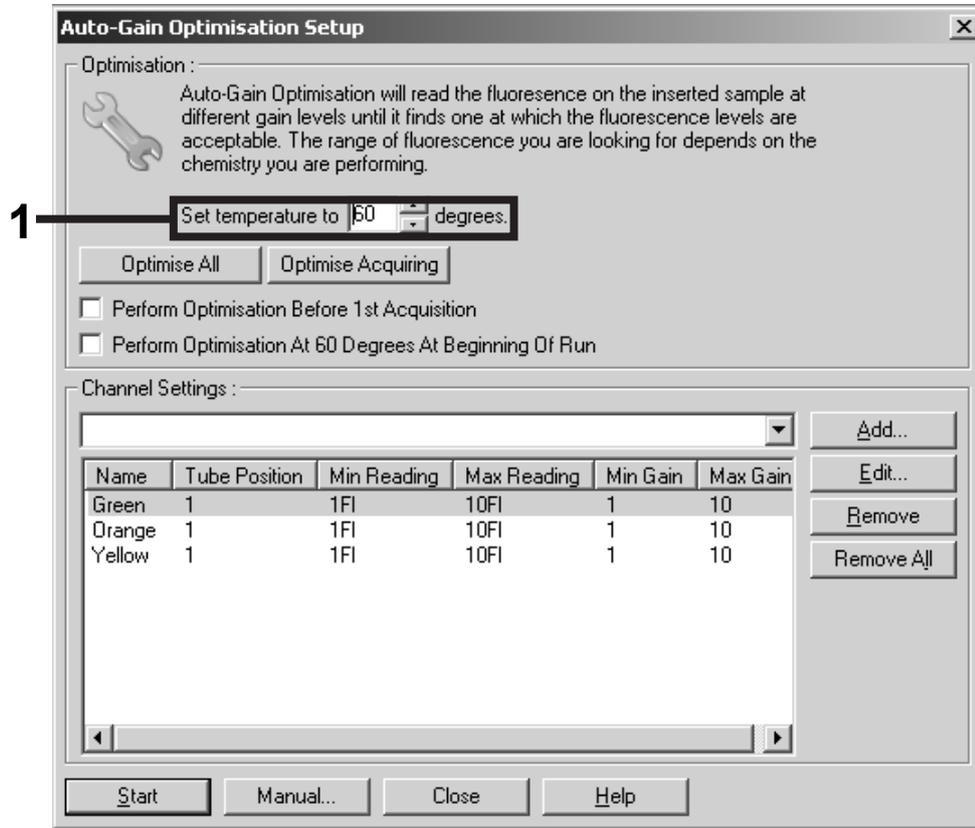


Figure 6. Adjusting the fluorescence channel sensitivity.

- Adjust the gain optimization. Choose a "Target Sample Range" from 1 FI up to 10 FI and an "Acceptable Gain Range" from 1 to 10 (Figure 7, example shown for channel Green). These adjustments need to be done for each channel (Green, Yellow, and Orange).

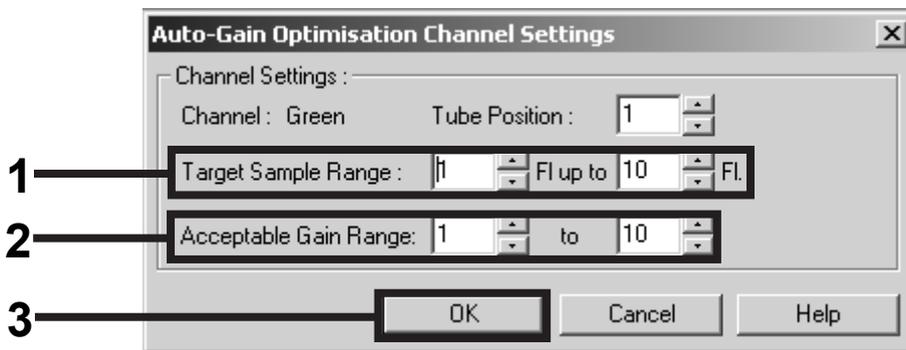


Figure 7. Adjusting the gain optimization for channel Green. These adjustments need to be done for each channel (Green, Yellow, and Orange).

10. Click the "Start" button to start gain optimization. After gain optimization is finished, click "Close" to return to the run wizard (Figure 8).

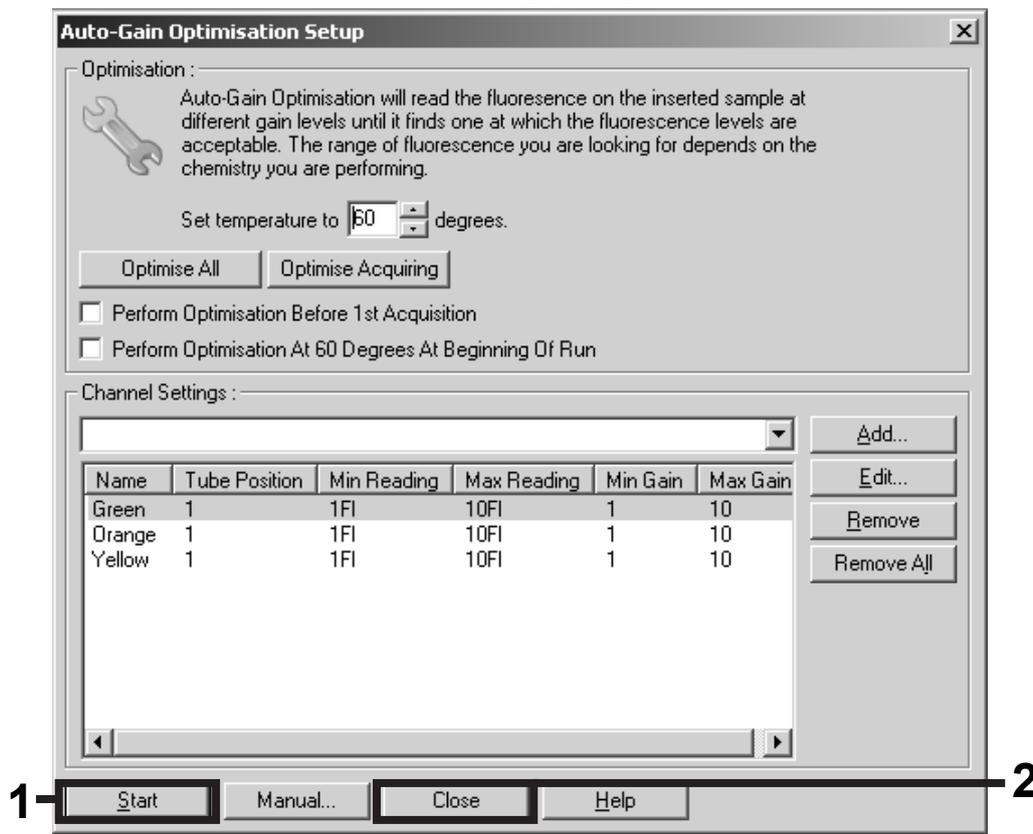


Figure 8. Gain optimization.

- The gain values determined by the channel calibration are saved automatically and are listed in the last menu window of the programming procedure (Figure 9). Click "Start Run".

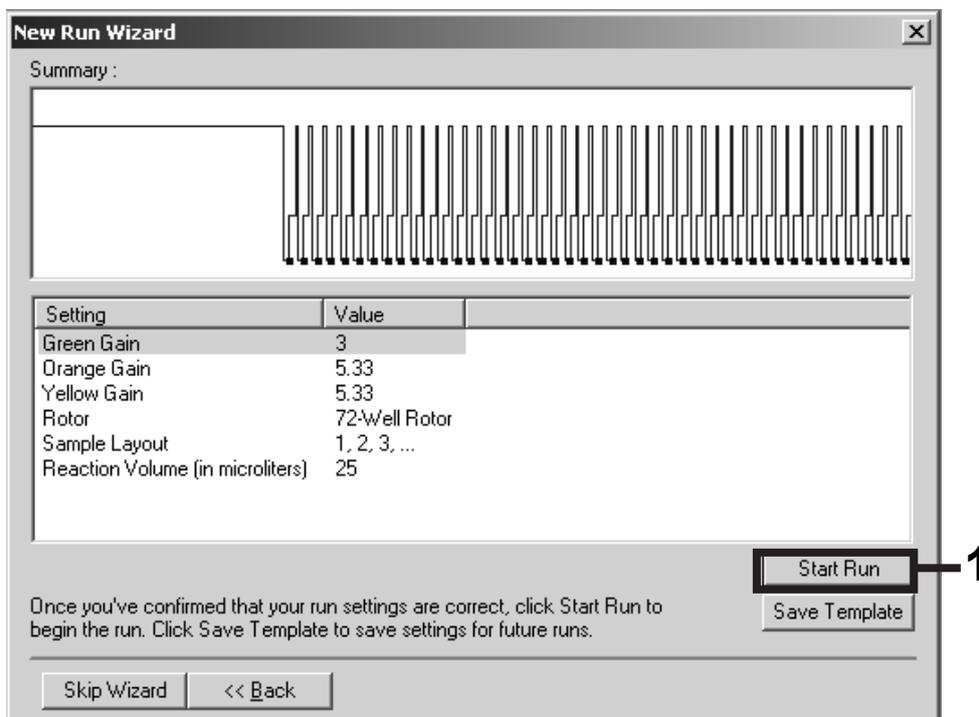


Figure 9. Starting the run.

- After starting the run, import the information from the cycler file by clicking the "Open" button, or edit the samples manually.
- For interpretation of results, see the instrument user manual and the relevant QIASymphony RGQ Application Sheet at www.qiagen.com/products/artusctngqsrqgkitce.aspx.

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